

# Folate

## Contents

- [Summary](#)
- [Function](#)
  - [One-carbon metabolism](#)
  - [Nutrient interactions](#)
- [Bioavailability](#)
- [Transport](#)
- [Deficiency](#)
  - [Causes](#)
  - [Symptoms](#)
- [The RDA](#)
  - [Determination](#)
  - [Dietary folate equivalents](#)
  - [Genetic variation](#)
- [Disease Prevention](#)
  - [Adverse pregnancy outcomes](#)
  - [Cardiovascular disease](#)
  - [Cancer](#)
  - [Alzheimer's disease and cognitive impairment](#)
- [Disease Treatment](#)
  - [Metabolic diseases](#)
- [Sources](#)
  - [Food](#)
  - [Supplements](#)
- [Safety](#)
  - [Toxicity](#)
  - [Drug interactions](#)
- [LPI Recommendation](#)
- [Authors and Reviewers](#)
- [References](#)

## [Español](#)

## Summary

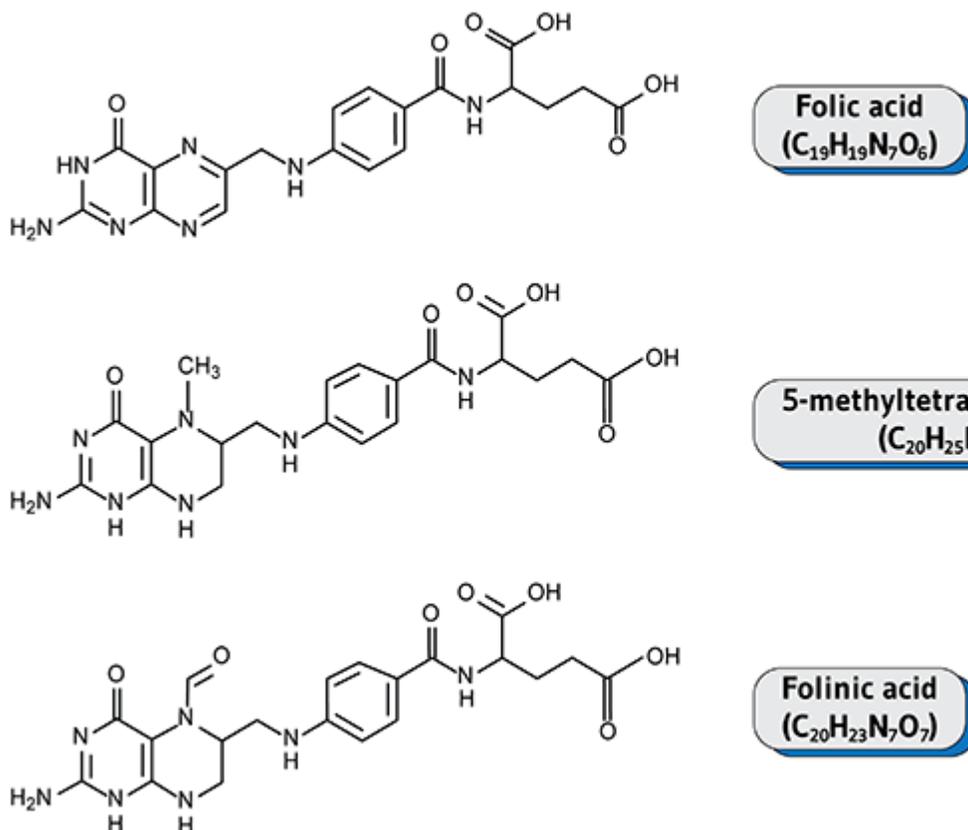
- Folate is a generic term referring to both natural folates in food and folic acid, the synthetic form used in [supplements](#) and [fortified](#) food. Folate is critical in the [metabolism](#) of [nucleic acid precursors](#) and several [amino acids](#), as well as in [methylation](#) reactions. ([More information](#)).
- Severe deficiency in either folate or [vitamin B<sub>12</sub>](#) can lead to [megaloblastic anemia](#), which causes fatigue, weakness, and shortness of breath. Improper treatment of vitamin B<sub>12</sub>-dependent megaloblastic anemia with high dose supplemental folic acid can potentially delay the diagnosis of vitamin B<sub>12</sub> deficiency and thus leave the individual at [risk](#) of developing irreversible brain damage. ([More information](#)).
- Folate status is influenced by the presence of genetic variations in folate metabolism, particularly those found in the 5,10-methylenetetrahydrofolate reductase (*MTHFR*) [gene](#). ([More information](#)).
- Inadequate folate status during early pregnancy increases the risk of [congenital anomalies](#). The introduction of mandatory folic acid [fortification](#) of refined grain products in the US in 1998 has reduced the prevalence of [neural tube defects](#) (NTDs) in newborns. Yet, folate status is considered inadequate in a majority of women of childbearing age worldwide. Moreover, genetic factors might modify the risk of NTDs by increasing the susceptibility to folate deficiency during pregnancy. Several studies are currently investigating the role of folic acid supplementation in the prevention of congenital anomalies other than NTDs. ([More information](#)).
- Folate deficiency and elevated concentrations of [homocysteine](#) in the blood are associated with increased risk

of [cardiovascular disease](#) (CVD). Although folic acid supplementation has been proven effective to control circulating homocysteine concentrations, the effect of homocysteine lowering on the incidence of CVD is still debated. ([More information](#)).

- Low folate status has been linked to increased [cancer](#) risk. However, [intervention trials](#) with high doses of folic acid have not generally shown any benefit on cancer incidence. ([More information](#)).
- [Prospective cohort studies](#) have reported an inverse association between folate status and [colorectal cancer](#) (CRC) risk, especially among men. The relationship between folate status and cancer risk is however complex and requires further research. ([More information](#)).
- Folate is essential for brain development and function. Low folate status and/or high homocysteine concentrations are associated with [cognitive](#) dysfunction in aging (from mild impairments to [dementia](#)). Whether supplemental B-vitamins, including folic acid, will have long-term benefits in maintaining cognitive health is not yet known. ([More information](#)).
- Several [autosomal recessive](#) disorders affecting folate transport and metabolism can be treated with high doses of folinic acid, a folate derivative. ([More information](#)).

Folate is a water-soluble B-vitamin, which is also known as vitamin B<sub>9</sub> or folacin. Naturally occurring folates exist in many chemical forms; folates are found in food, as well as in metabolically active forms in the human body. Folic acid is the major synthetic form found in fortified foods and vitamin supplements. Other synthetic forms include folinic acid (**Figure 1**) and levomefolic acid. Folic acid has no biological activity unless converted into folates (**1**). In the following discussion, forms found in food or the body are referred to as "folates," while the form found in supplements or fortified food is referred to as "folic acid."

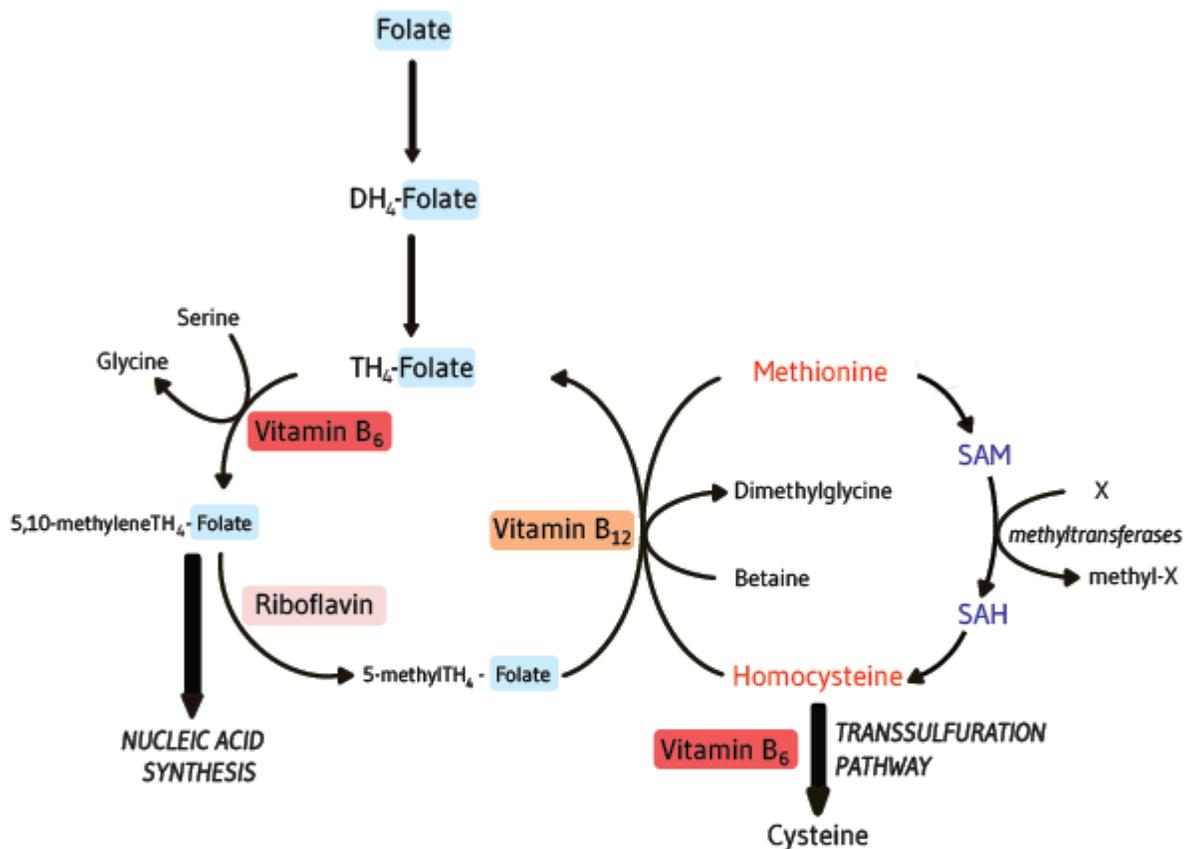
**Figure 1. Chemical Structures**



## Function

### One-carbon metabolism

The only function of folate [coenzymes](#) in the body appears to be in mediating the transfer of [one-carbon units](#) (**2**). Folate coenzymes act as acceptors and donors of one-carbon units in a variety of reactions critical to the [metabolism](#) of [nucleic acids](#) and [amino acids](#) (**Figure 2**) (**3**).

**Figure 2. Overview of One-carbon Metabolism**

5,10-methylenetetrahydrofolate is required for the synthesis of nucleic acids, and 5-methyltetrahydrofolate is required for the formation of methionine from homocysteine. Methionine, in the form of methyl donor S-adenosylmethionine (SAM), is essential to many biological methylation reactions, including DNA methylation. Methylenetetrahydrofolate reductase (MTHFR) is a riboflavin (FAD)-dependent enzyme that catalyzes the reduction of 5,10-methylenetetrahydrofolate to 5-methyltetrahydrofolate. SAM, S-adenosylmethionine; SAH, S-adenosylhomocysteine; TH<sub>4</sub>-Folate, Tetrahydrofolate.

### *Nucleic acid metabolism*

Folate [coenzymes](#) play a vital role in [DNA metabolism](#) through two different pathways. (1) The [synthesis](#) of DNA from its [precursors](#) (thymidine and purines) is dependent on folate coenzymes. (2) A folate coenzyme is required for the synthesis of [methionine](#) from [homocysteine](#), and methionine is required for the synthesis of S-adenosylmethionine (SAM). SAM is a methyl group (one-carbon unit) donor used in most biological [methylation](#) reactions, including the methylation of a number of sites within DNA, [RNA](#), [proteins](#), and [phospholipids](#). The methylation of DNA plays a role in controlling [gene expression](#) and is critical during cell [differentiation](#). Aberrations in DNA methylation have been linked to the development of [cancer](#) (see [Cancer](#)).

### *Amino acid metabolism*

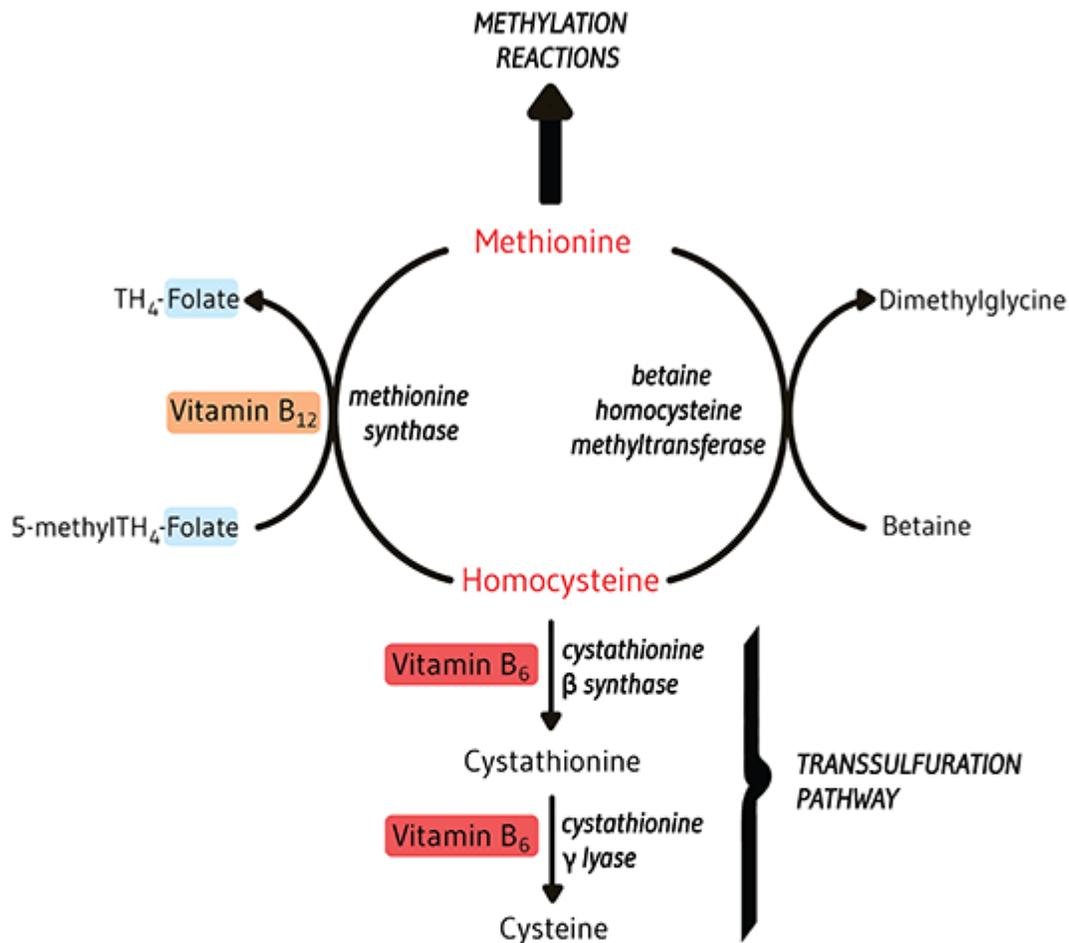
Folate [coenzymes](#) are required for the [metabolism](#) of several important [amino acids](#), namely [methionine](#), cysteine, serine, glycine, and histidine. The [synthesis](#) of methionine from [homocysteine](#) is catalyzed by methionine synthase, an enzyme that requires not only folate (as 5-methyltetrahydrofolate) but also [vitamin](#)

[B<sub>12</sub>](#). Thus, folate (and/or vitamin B<sub>12</sub>) deficiency can result in decreased synthesis of methionine and an accumulation of homocysteine. Elevated blood concentrations of homocysteine have been considered for many years to be a [risk](#) factor for some chronic diseases, including [cardiovascular disease](#) and [dementia](#) (see [Disease Prevention](#)).

## Nutrient interactions

### *Vitamin B<sub>12</sub> and vitamin B<sub>6</sub>*

The [metabolism](#) of [homocysteine](#), an intermediate in the metabolism of sulfur-containing [amino acids](#), provides an example of the interrelationships among nutrients necessary for optimal physiological function and health. Healthy individuals utilize two different pathways to metabolize homocysteine (**Figure 3**). One pathway (methionine synthase) [synthesizes methionine](#) from homocysteine and is dependent on both folate and [vitamin B<sub>12</sub>](#) as [cofactors](#). The other pathway converts homocysteine to another amino acid, cysteine, and requires two [vitamin B<sub>6</sub>](#)-dependent [enzymes](#). Thus, the concentration of homocysteine in the blood is regulated by three B-vitamins: folate, vitamin B<sub>12</sub>, and vitamin B<sub>6</sub> ([4](#)). In some individuals, riboflavin (vitamin B<sub>2</sub>) is also involved in the regulation of homocysteine concentrations (see the article on [Riboflavin](#)).

**Figure 3. Homocysteine Metabolism**

Homocysteine is methylated to form the essential amino acid methionine in two pathways. The reaction of homocysteine remethylation catalyzed by the vitamin B<sub>12</sub>-dependent methionine synthase captures a methyl group from the folate-dependent one-carbon pool (5-methyltetrahydrofolate). A second pathway requires betaine (N,N,N-trimethylglycine) as a methyl donor for the methylation of homocysteine catalyzed by betaine homocysteine methyltransferase. The catabolic pathway of homocysteine, known as the transsulfuration pathway, converts homocysteine to the amino acid cysteine via two vitamin B<sub>6</sub> (PLP)-dependent enzymes. Cystathionine β synthase catalyzes the condensation of homocysteine with serine to form cystathionine, and cystathionine is then converted to cysteine, α-ketobutyrate, and ammonia by cystathionine γ lyase. TH<sub>4</sub>-Folate, Tetrahydrofolate.

### Riboflavin

Although less well recognized, folate has an important metabolic interaction with [riboflavin](#). Riboflavin is a [precursor](#) of flavin adenine dinucleotide (FAD), a [coenzyme](#) required for the activity of the folate-metabolizing [enzyme](#), 5,10-methylenetetrahydrofolate reductase (MTHFR). FAD-dependent MTHFR in turn [catalyzes](#) the reaction that generates 5-methyltetrahydrofolate (see [Figure 2](#) above). This active form of folate is required to form [methionine](#) from [homocysteine](#). Along with other B-vitamins, higher riboflavin intakes have been associated with decreased [plasma](#) homocysteine concentrations ([5](#)). The effects of riboflavin on folate [metabolism](#) appear to be greatest in individuals [homozygous](#) for the common c.677C>T [polymorphism](#) (i.e., TT genotype) in the *MTHFR* [gene](#) (see [Genetic variation in folate requirements](#)) ([6](#)). These individuals (about 10% of adults worldwide) typically present with low folate status, along with elevated homocysteine

concentrations, particularly when folate and/or riboflavin intake is suboptimal. The elevated homocysteine concentration in these individuals, however, is highly responsive to lowering with riboflavin supplementation, confirming the importance of the riboflavin-*MTHFR* interaction (7).

### *Vitamin C*

[Vitamin C](#) may limit degradation of natural folate [coenzymes](#) and [supplemental](#) folic acid in the stomach and thus improve folate [bioavailability](#). A [cross-over trial](#) in nine healthy men found that oral co-administration of 5-methyltetrahydrofolic acid (343 µg) and vitamin C (289 mg or 974 mg) was associated with higher concentrations of serum folate compared to 5-methyltetrahydrofolic acid alone (8). Moreover, a recent study suggested that several genetic variations of folate [metabolism](#) might influence the effect of vitamin C on folate metabolism (9).

## Bioavailability

Dietary folates exist predominantly in the polyglutamyl form (containing several [glutamate](#) residues), whereas folic acid—the synthetic vitamin form—is a monoglutamate, containing just one glutamate moiety. In addition, natural folates are reduced molecules, whereas folic acid is fully oxidized. These chemical differences have major implications for the [bioavailability](#) of the [vitamin](#) such that folic acid is considerably more bioavailable than naturally occurring food folates at equivalent intake levels.

The [intestinal](#) absorption of dietary folates is a two-step process that involves the [hydrolysis](#) of folate polyglutamates to the corresponding monoglutamyl derivatives, followed by their transport into intestinal cells. There, folic acid is converted into a naturally occurring folate, namely 5-methyltetrahydrofolate, which is the major circulating form of folate in the human body (see [Figure 1](#) above).

The bioavailability of naturally occurring folates is inherently limited and variable. There is much variability in the ease with which folates are released from different food matrices, and the polyglutamyl "tail" is removed (de-conjugation) before uptake by intestinal cells. Also, other dietary constituents can contribute to instability of labile folates during the processes of digestion. As a result, naturally occurring folates show incomplete bioavailability compared with folic acid. The bioavailability of folic acid, in contrast, is assumed to be 100% when ingested as a [supplement](#), while folic acid in [fortified](#) food is estimated to have about 85% the bioavailability of supplemental folic acid.

Of note, folate recommendations in the US and certain other countries are now expressed as Dietary Folate Equivalents (DFEs), a calculation that was devised to take into account the greater bioavailability of folic acid compared to naturally occurring dietary folates (see [The Recommended Dietary Allowance](#)).

## Transport

Folate and its [coenzymes](#) require transporters to cross [cell membranes](#). Folate transporters include the reduced folate carrier (RFC), the proton-coupled folate transporter (PCFT), and the folate receptor proteins, FR $\alpha$  and FR $\beta$ . Folate [homeostasis](#) is supported by the ubiquitous distribution of folate transporters, although abundance and importance vary among tissues (10). PCFT plays a major role in folate intestinal transport since [mutations](#) affecting the [gene](#) encoding PCFT cause hereditary folate [malabsorption](#). Defective PCFT also leads to impaired folate transport into the brain (see [Disease Treatment](#)). FR $\alpha$  and RFC are also critical for folate transport across the blood-brain barrier when extracellular folate is either low or high, respectively. Folate is essential for the proper development of the embryo and the fetus. The [placenta](#) is known to concentrate folate to the fetal circulation, leading to higher folate concentrations in the fetus compared to those found in the pregnant woman. All three types of [receptors](#) have been associated with folate transport across the placenta during pregnancy (11).

## Deficiency

## Causes

Folate deficiency is most often caused by a dietary insufficiency; however, folate deficiency can also occur in a number of other situations. For example, chronic and heavy alcohol consumption is associated with diminished absorption of folate (in addition to low dietary intake), which can lead to folate deficiency (12). Smoking is also associated with low folate status. In one study, folate concentrations in blood were about 15% lower in smokers compared to nonsmokers (13). Additionally, impaired folate transport to the fetus has been described in pregnant women who either smoked or abused alcohol during their pregnancy (14, 15).

Pregnancy is a time when the folate requirement is greatly increased to sustain the demand for rapid cell replication and growth of fetal, placental, and maternal tissue. Conditions such as cancer or inflammation can also result in increased rates of cell division and metabolism, causing an increase in the body's demand for folate (16). Moreover, folate deficiency can result from some malabsorptive conditions, including inflammatory bowel diseases (Crohn's disease and ulcerative colitis) and celiac disease (17). Several medications may also contribute to folate deficiency (see Drug interactions). Finally, a number of genetic diseases affecting folate absorption, transport, or metabolism can cause folate deficiency or impede its metabolic functions (see Disease Treatment).

## Symptoms

Clinical folate deficiency leads to megaloblastic anemia, which is reversible with folic acid treatment. Rapidly dividing cells like those derived from bone marrow are most vulnerable to the effects of folate deficiency since DNA synthesis and cell division are dependent on folate coenzymes. When folate supply to the rapidly dividing cells of the bone marrow is inadequate, blood cell division is reduced, resulting in fewer but larger red blood cells. This type of anemia is called megaloblastic or macrocytic anemia, referring to the enlarged, immature red blood cells. Neutrophils, a type of white blood cell, become hypersegmented, a change that can be found by examining a blood sample microscopically. Because normal red blood cells have a lifetime in the circulation of approximately four months, it can take months for folate-deficient individuals to develop the characteristic megaloblastic anemia. Progression of such an anemia leads to a decreased oxygen carrying capacity of the blood and may ultimately result in symptoms of fatigue, weakness, and shortness of breath (1). It is important to point out that megaloblastic anemia resulting from folate deficiency is identical to the megaloblastic anemia resulting from vitamin B<sub>12</sub> deficiency, and further clinical testing is required to diagnose the true cause of megaloblastic anemia (see Toxicity).

Individuals in the early stages of folate deficiency may not show obvious symptoms, but blood concentrations of homocysteine may increase (see Disease Prevention). Yet, the concentration of circulating homocysteine is not a specific indicator of folate status, as elevated homocysteine can be the result of vitamin B<sub>12</sub> and other B-vitamin deficiencies, lifestyle factors, and renal insufficiency. Subclinical deficiency is typically detected by measurement of folate concentrations in serum/plasma or in red blood cells.

## The Recommended Dietary Allowance (RDA)

### Determination of the RDA

Traditionally, the dietary folate requirement was defined as the amount needed to prevent a deficiency severe enough to cause symptoms like anemia. The most recent RDA (1998; Table 1) was based primarily on the adequacy of red blood cell folate concentrations at different levels of folate intake, as judged by the absence of abnormal hematological indicators. Red cell folate has been shown to correlate with liver folate stores and is used as an indicator of long-term folate status. Plasma folate reflects recent folate intake and is not a reliable biomarker for folate status. Maintenance of normal blood homocysteine concentrations, an indicator of one-carbon metabolism, was considered only as an ancillary indicator of adequate folate intake.

Because pregnancy is associated with a significant increase in cell division and other metabolic processes that require folate [coenzymes](#), the RDA for pregnant women is considerably higher than for women who are not pregnant (3). However, the prevention of [neural tube defects](#) (NTDs) was not considered when setting the RDA for pregnant women. Rather, reducing the [risk](#) of NTDs was considered in a separate recommendation for women capable of becoming pregnant (see [Disease Prevention](#)), because the crucial events in the development of the neural tube occur before many women are aware that they are pregnant (18).

### Dietary Folate Equivalents (DFEs)

When the Food and Nutrition Board of the US Institute of Medicine set the new dietary recommendation for folate, they introduced a new unit, the Dietary Folate Equivalent (DFE) (1). Use of the DFE reflects the higher [bioavailability](#) of synthetic folic acid found in [supplements](#) and [fortified](#) food compared to that of naturally occurring food folates (18).

- 1 microgram ( $\mu\text{g}$ ) of food folate provides 1  $\mu\text{g}$  of DFEs
- 1  $\mu\text{g}$  of folic acid taken with meals or as fortified food provides 1.7  $\mu\text{g}$  of DFEs
- 1  $\mu\text{g}$  of folic acid (supplement) taken on an empty stomach provides 2  $\mu\text{g}$  of DFEs

For example, a serving of food containing 60  $\mu\text{g}$  of folate would provide 60  $\mu\text{g}$  of DFEs, while a serving of pasta fortified with 60  $\mu\text{g}$  of folic acid would provide  $1.7 \times 60 = 102$   $\mu\text{g}$  of DFEs due to the higher bioavailability of folic acid. A folic acid supplement of 400  $\mu\text{g}$  taken on an empty stomach would provide 800  $\mu\text{g}$  of DFEs. It should be noted that DFEs were determined in studies with adults and whether folic acid in infant formula is more bioavailable than folates in mother's milk has not been studied. Use of DFEs to determine a folate requirement for the infant would not be desirable.

Table 1. Recommended Dietary Allowance for Folate in Dietary Folate Equivalents (DFEs)

Life Stage	Age	Males ( $\mu\text{g/day}$ )	Females ( $\mu\text{g/day}$ )
Infants	0-6 months	65 (AI)	65 (AI)
Infants	7-12 months	80 (AI)	80 (AI)
Children	1-3 years	150	150
Children	4-8 years	200	200
Children	9-13 years	300	300
Adolescents	14-18 years	400	400
Adults	19 years and older	400	400
Pregnancy	all ages	-	600
Breast-feeding	all ages	-	500

### Genetic variation in folate requirements

A common [polymorphism](#) or variation in the sequence of the [gene](#) for the [enzyme](#), 5, 10-methylenetetrahydrofolate reductase (MTHFR), known as the *MTHFR* c.677C>T polymorphism, results in a thermolabile enzyme (19). The substitution of a cytosine (C) by a thymine (T) at [nucleotide](#) 677 in the exon 4 of *MTHFR* gene leads to an alanine-to-valine transition in the catalytic domain of the enzyme. Depending on the population, 20% to 53% of individuals may have inherited one T copy (677C/T genotype), and 3% to 32% of individuals may have inherited two T copies (677T/T genotype) for the *MTHFR* gene (20). MTHFR [catalyzes](#) the reduction of 5,10-methylenetetrahydrofolate (5,10-methylene THF) into 5-methyl

tetrahydrofolate (5-MeTHF). The latter is the folate [coenzyme](#) required to form [methionine](#) from [homocysteine](#) (see [Figure 2](#) above). MTHFR activity is greatly diminished in heterozygous 677C/T (-30%) and homozygous 677T/T (-65%) individuals compared to those with the 677C/C genotype ([21](#)). Homozygosity for the [mutation](#) (677T/T) is linked to lower concentrations of folate in red blood cells and higher blood concentrations of homocysteine ([22, 23](#)). Improving folate nutritional [status](#) in elderly women with the T allele reduced [plasma](#) homocysteine concentration ([24](#)). An important unanswered question about folate is whether the present [RDA](#) is enough to compensate for the reduced MTHFR enzyme activity in individuals with at least one T allele, or whether those individuals have a higher folate requirement than the RDA ([25](#)).

## Disease Prevention

### Adverse pregnancy outcomes

#### *Neural tube defects*

Fetal growth and development are characterized by widespread cell division. Adequate folate is critical for [DNA](#) and [RNA synthesis](#). [Neural tube defects](#) (NTDs) arise from failure of embryonic neural tube closure between the 21<sup>st</sup> and 27<sup>th</sup> days after conception, a time when many women may not even realize they are pregnant ([26](#)). NTDs include various malformations, such as lesions of the brain (e.g., [anencephaly](#), encephalocele) or lesions of the spine ([spina bifida](#)), which are devastating and life-threatening ([27](#)). The prevalence of NTDs in the United States prior to [fortification](#) of food with folic acid was estimated to be 1 per 1,000 pregnancies ([1](#)). Results of [randomized trials](#) have demonstrated 60% to 100% reductions in NTD cases when women consumed folic acid [supplements](#) in addition to a varied diet during the periconceptional period (about one month before and at least one month after conception) ([28, 29](#)). The results of these and other studies prompted the US Public Health Service to recommend that all women capable of becoming pregnant consume 400 µg of folic acid daily to prevent NTDs. Women with a previously affected pregnancy were also advised to receive 4,000 µg (4 mg) of folic acid daily in order to reduce NTD recurrence ([30](#)). These recommendation were made to all women of childbearing age because adequate folate must be available very early in pregnancy, and because many pregnancies in the US are unplanned ([31](#)).

Despite the effectiveness of folic acid supplementation in improving folate status, it appears that globally only 30% of women who become pregnant correctly follow the recommendation, and there is some concern that young women from minority ethnic groups and lower socio-economic backgrounds are the least likely to follow the recommendation ([32-34](#)). To decrease the incidence of NTDs, the US FDA implemented legislation in 1998 requiring the fortification of all enriched grain products with 1.4 mg of folic acid per kg of grain (see [Sources](#)). The required level of folic acid fortification in the US was initially estimated to provide 100 µg of additional folic acid in the average person's diet, though it probably provides even more due to overuse of folic acid by food manufacturers ([25, 35](#)). The National Birth Defects Prevention Network reported about a 30% decrease in the prevalence of NTDs in the US compared to the pre-fortification period, and the post-fortification prevalence of NTDs is 0.69 cases per 1,000 live births and fetal deaths ([36](#)).

Also, a genetic component in NTD [etiology](#) is evidenced by the increased [risk](#) in women with a family history of an NTD and also by variations in risk among ethnicities ([37](#)). Moreover, NTD occurrence can be attributed to specific folate-[gene](#) interactions. The *MTHFR* c.677C>T [polymorphism](#) and other genetic variations can increase the folate requirement and susceptibility for an NTD-affected pregnancy. Prior to the fortification era, a [case-control study](#) showed that both red blood cell and [serum](#) folate concentrations were significantly lower in pregnant women with the T/T and C/T variants compared to the wild-type C/C genotype ([22](#)), suggesting inadequate folate [metabolism](#) with specific maternal genotypes. A [meta-analysis](#) of 25 case-control studies, including 2,429 case mothers and 3,570 control mothers, showed a positive association between the maternal *MTHFR* c.677C>T polymorphism and NTDs ([38](#)). Another *MTHFR* variant, an A-to-C change at position 1298, has also been associated with reduced MTHFR activity and increased NTD risk ([39](#)). Individuals [heterozygous](#) for both of these *MTHFR* variants (677C/T + 1298A/C) exhibit lower [plasma](#) folate and higher

[homocysteine](#) concentrations than individuals with 677C/T + 1298A/A (40). Combined genotypes with [homozygosity](#) G/G for the reduced folate carrier transporter (*RFC-1*) polymorphism (c.80A>G) could further contribute to NTD occurrence (41). The degree of NTD risk was also assessed with additional *MTHFR* polymorphisms (c.116C>T, c.1793G>A) (42), as well as with [mutations](#) affecting other [enzymes](#) of the [one-carbon](#) metabolism, including methionine synthase (*MTR* c.2756A>G) (43), methionine synthase reductase (*MTRR* c.66A>G) (44), and methylenetetrahydrofolate dehydrogenase (*MTHFD1* c.1958G>A) (45). While maternal genotype can impact pregnancy outcome, it appears that gene-gene interactions between mother and fetus influence it further. The risk of NTD was increased by certain genetic combinations, including maternal (*MTHFR* c.677C>T)-fetal (*MTHFR* c.677C>T) and maternal (*MTRR* c.66A>G)-fetal (*MTHFR* c.677C>T) interactions (43, 44, 46). Finally, [vitamin B<sub>12</sub>](#) status has been associated with NTD risk modification in the presence of specific polymorphisms in one-carbon metabolism (47).

### *Cardiovascular malformations*

[Congenital anomalies](#) of the heart are a major cause of infant mortality but also cause deaths in adulthood (48). Using data from the European Registration of Congenital Anomalies and Twins (EUROCAT) database, a [case-control study](#), involving 596 cases and 2,359 controls, found that consumption of at least 400 µg/day of folic acid during the periconceptual period (one month before conception through eight weeks' post-conception, covering the period of embryonic heart development) was associated with an 18% reduced [risk](#) of congenital heart defects (49). Recent meta-analyses of 20 to 25 case-control and family-based studies observed positive associations between maternal, fetal, or paternal *MTHFR* c.677C>T variant and incidence of congenital heart defects (50, 51). Additional studies are needed to elucidate the effects of [gene-nutrient](#) interactions on the risk of congenital heart defects; however, the currently available research indicates that adequate folate intake may play an important role.

### *Orofacial clefts*

Maternal folate status during pregnancy may influence the [risk](#) of [congenital anomalies](#) called orofacial clefts, namely cleft lip with or without cleft palate (CL/P) (52). A population-based [case-control study](#) in Norway investigated the impact of folic acid [supplements](#) in mothers of 377 newborns with CL/P, 196 with cleft palate only (CPO) and 763 controls (53). Although dietary intakes or supplements (during the first three months of pregnancy) on their own did not significantly modify the risk of CL/P, the study reported a 64% lower risk among women taking multivitamin and folic acid (≥400 µg daily) supplements in addition to dietary folates. In the same population, [polymorphisms](#) in the cystathionine β-synthase (*CBS*) [gene](#) (c.699C>T) or *MTHFR* gene (c.677C>T; when folate intake was below 400 µg/day) appeared protective, while other gene variants in the folate/[one-carbon](#) metabolism could not be linked to CL/P (54, 55). However, a recent [meta-analysis](#) of 18 studies showed an elevation of CL/P risk with the maternal 677T/T [homozygosity](#) (56). Additional studies are needed to evaluate the risk of CL/P while integrating both genetic polymorphism and folate intake parameters. [Epidemiological](#) evidence supporting a role for folate in the risk of CPO is lacking.

### *Other adverse pregnancy outcomes*

Low birth weight has been associated with increased [risk](#) of mortality during the first year of life and may also influence health outcomes during adulthood (57). A recent [systematic review](#) and [meta-analysis](#) of eight [randomized controlled trials](#) found a positive association between folic acid [supplementation](#) and birth weight; no association with length of [gestation](#) was observed (58). Additionally, a [prospective cohort study](#) of 306 pregnant adolescents associated low folate intakes and maternal folate status during the third trimester of pregnancy with higher incidence of small for gestational age births (birth weight <10<sup>th</sup> percentile) (59). Moreover, the maternal c.677C>T *MTHFR* genotype and increased [homocysteine](#) concentrations, considered an indicator of functional folate deficiency, have been linked to lower birth weights (60).

Elevated blood homocysteine concentrations have also been associated with increased incidence of miscarriage and other pregnancy complications, including [preeclampsia](#) and [placental abruption](#) (61). A large

[retrospective study](#) showed that [plasma](#) homocysteine in Norwegian women was strongly related to adverse outcomes and complications, including preeclampsia, premature delivery, and very low birth weight, in previous pregnancies (62). A recent meta-analysis of 51 prospective cohort studies linked the c.677C>T *MTHFR* variant with increased risk of preeclampsia in Caucasian and East Asian populations, reinforcing the notion that folate [metabolism](#) may play a role in the condition (63). A large multicenter, randomized, controlled trial, the Folic Acid Clinical Trial (FACT), has been initiated to evaluate whether the daily supplementation of up to 5.1 mg of folic acid throughout pregnancy could prevent preeclampsia and other adverse outcomes (e.g., maternal death, placental abruption, preterm delivery) in high-risk women (64). Adequate folate intake during pregnancy protects against [megaloblastic anemia](#) (65). A recent [case-control study](#) found a reduction in risk of autism spectrum disorders with daily folic acid consumption of 600 µg or more before and during pregnancy when mother and child carried the c.677C>T *MTHFR* genotype (66).

Thus, it is reasonable to maintain folic acid supplementation throughout pregnancy, even after closure of the neural tube, in order to decrease the risk of other problems during pregnancy. Moreover, recent systematic reviews of [observational studies](#) found no evidence of an association between folate exposure during pregnancy and adverse health outcomes in offspring, in particular childhood [asthma](#) and allergies (67, 68).

## Cardiovascular disease

### *Homocysteine and cardiovascular disease*

The results of more than 80 studies indicate that even moderately elevated concentrations of [homocysteine](#) in the blood increase the [risk](#) of [cardiovascular disease](#) (CVD) (4). Possible predispositions to vascular accidents have also been linked to genetic deficiencies in homocysteine [metabolism](#) in certain populations (69). The mechanism by which homocysteine may increase the risk of vascular disease has been the subject of a great deal of research, but it may involve adverse effects of homocysteine on blood clotting, arterial [vasodilation](#), and thickening of arterial walls (70). Although increased homocysteine concentrations in the blood have been consistently associated with increased risk of CVD, it is unclear whether lowering circulating homocysteine will reduce CVD risk (see [Folate and homocysteine](#)). Research had initially predicted that a prolonged decrease in [serum](#) homocysteine level of 3 micromoles/liter would lower the risk of CVD by up to 25% and be a reasonable treatment goal for individuals at high risk (71, 72). However, the analysis of recent [clinical trials](#) of B-vitamin supplementation has shown that lowering homocysteine concentrations did not prevent the occurrence of a second [cardiovascular](#) event in patients with existing CVD (73, 74). Consequently, the American Heart Association recommends screening for elevated total homocysteine concentrations only in "high risk" individuals, for example, in those with personal or family history of premature cardiovascular disease, malnutrition or [malabsorption syndromes](#), [hypothyroidism](#), kidney failure, [lupus](#), or individuals taking certain medications (nicotinic acid, theophylline, [bile acid](#)-binding resins, methotrexate, and L-dopa).

### *Folate and homocysteine*

Folate-rich diets have been associated with decreased [risk](#) of [CVD](#), including [coronary artery disease](#), [myocardial infarction](#) (heart attack), and [stroke](#). A study that followed 1,980 Finnish men for 10 years found that those who consumed the most dietary folate had a 55% lower risk of an acute coronary event when compared to those who consumed the least dietary folate (75). Of the three B-vitamins that regulate [homocysteine](#) concentrations, folic acid has been shown to have the greatest effect in lowering basal concentrations of homocysteine in the blood when there is no coexisting deficiency of [vitamin B<sub>12</sub>](#) or [vitamin B<sub>6</sub>](#) (see [Nutrient interactions](#)) (76). Increasing folate intake through folate-rich food or [supplements](#) has been found to reduce homocysteine concentrations (77). Besides, blood homocysteine concentrations have declined since the FDA mandated folic acid [fortification](#) of the grain supply in the US (25). A [meta-analysis](#) of 25 [randomized controlled trials](#), including almost 3,000 subjects, found that folic acid supplementation with 800 µg/day or more could achieve a maximal 25% reduction in [plasma](#) homocysteine concentrations. In this meta-analysis, daily doses of 200 µg and 400 µg of folic acid were associated with a 13% and 20% reduction in plasma homocysteine, respectively (78). A supplement regimen of 400 µg of folic acid, 2 mg of vitamin B<sub>6</sub>,

and 6 µg of vitamin B<sub>12</sub> has been advocated by the American Heart Association if an initial trial of a folate-rich diet (see [Sources](#)) is not successful in adequately lowering homocysteine concentrations (79).

Several [polymorphisms](#) in folate/[one-carbon](#) metabolism modify homocysteine concentrations in blood (80). In particular, the effect of the c.677C>T *MTHFR* variant has been examined in relation to folic acid fortification policies worldwide. The analysis of [randomized](#) trials, including 59,995 subjects without a history of CVD, revealed that the difference in homocysteine concentrations between T/T and C/C genotypes was greater in low-folate regions compared to regions with food fortification policy (3.12 vs. 0.13 micromoles/liter) (81). Although folic acid supplementation effectively decreases homocysteine concentrations, it is not yet clear whether it also decreases risk for CVD. A recent meta-analysis of 19 randomized [clinical trials](#), including 47,921 subjects with preexisting cardiovascular or [renal](#) disease, found that homocysteine lowering through folic acid and other B-vitamin supplementation failed to reduce the incidence of CVD despite significant reductions in plasma homocysteine concentrations (74). Other meta-analyses have confirmed the lack of causality between the lowering of homocysteine and the risk of CVD (80-82), including the risk of stroke (83, 84). Consequently, the American Heart Association removed its recommendation for using folic acid to prevent cardiovascular disease in high-risk women (85). It should be noted that the majority of prevention trials to date have been performed in CVD patients with advanced disease. The evidence supporting a beneficial role for folate and related B-vitamins appears to be strongest for the primary prevention of stroke (86). The introduction of mandatory folic acid fortification has been associated with a decline in stroke-related mortality in North America, adding further support to the potential benefit of enhancing folate status and/or lowering homocysteine in the prevention of stroke (87).

Despite the controversy regarding the role of homocysteine lowering in CVD prevention, some studies have investigated the effect of folic acid supplementation on the development of [atherosclerosis](#), a known risk factor for vascular accidents. The measurement of the [carotid](#) intima-media thickness (CIMT) is a surrogate endpoint for early atherosclerosis and a predictor for [cardiovascular](#) events (88). The meta-analysis of 10 randomized trials testing the effect of folic acid supplementation showed a significant reduction in CIMT in subjects with chronic kidney diseases and in those at risk for CVD, but not in healthy participants (89). [Endothelial](#) dysfunction is a common feature in atherosclerosis and vascular disease. High doses of folic acid (400-10,000 µg/day) have been associated with improvements in vascular health in both healthy and CVD subjects (90). Although recent trials failed to demonstrate any cardiovascular protection from folic acid supplementation, low folate intake is a known risk factor for vascular disease, and more research is needed to explore the role of folate in maintaining vascular health (91).

## Cancer

[Cancer](#) is thought to arise from [DNA](#) damage in excess of ongoing DNA repair and/or the inappropriate expression of critical [genes](#). Because of the important roles played by folate in DNA and [RNA synthesis](#) and [methylation](#), it is possible that inadequate folate intake contributes to genome instability and [chromosome](#) breakage that often characterize cancer development. In particular, DNA replication and repair are critical for genome maintenance, and the shortage in [nucleotides](#) caused by folate deficiency might lead to genome instability and DNA [mutations](#). A decrease in 5,10-methylene THF can compromise the conversion of deoxyuridine monophosphate (dUMP) to deoxythymidine monophosphate (dTMP) by the [enzyme](#) thymidylate synthase (TS), causing uracil accumulation and thymine depletion. This could then lead to uracil misincorporation into DNA during replication or repair, and cause DNA damage, including point mutations and strand breaks (92). Since 5,10-methylene THF is also the MTHFR enzyme substrate, it is plausible that a reduction of MTHFR activity with the c.677C>T polymorphism may increase the use of 5,10-methylene THF for thymidylate synthesis and prevent DNA damage. However, this hypothesis might only be valid in a situation of folate deficiency (93). Conversely, it was argued that folic acid supplementation could fuel DNA synthesis, therefore promoting tumor growth. This is supported by the observation that TS can function like a tumor promoter (oncogene), while a reduction in TS activity is linked to a lower [risk](#) of cancer (94, 95). Additionally, antifolate molecules that block the thymidylate synthesis pathway are successfully used in cancer therapy (96). Folate also controls the homocysteine/[methionine](#) cycle and the pool of S-adenosylmethionine (SAM), the methyl donor for methylation reactions. Thus, folate deficiency may impair

DNA and [protein](#) methylation and alter the [expression](#) of genes involved in DNA repair, [proliferation](#) and cell death. Global DNA hypomethylation, a typical hallmark of cancer, causes genome instability and chromosome breaks (reviewed in [97](#)).

The consumption of at least five servings of fruit and vegetables daily has been consistently associated with a decreased incidence of cancer ([98](#)). Fruit and vegetables are excellent sources of folate, which may play a role in their anti-carcinogenic effect. [Observational studies](#) have found diminished folate [status](#) to be associated with site-specific cancers. While food [fortification](#) is mandatory in the US (since 1998; see [Sources](#)), concerns about the impact of high folic acid intakes on health have delayed the practice in several other countries ([99](#)). However, the most recent [meta-analyses](#) of folic acid [intervention trials](#) (supplemental doses ranging from 500 to 5,000 µg/day for at least one year) did not show any specific benefit or harm regarding total and site-specific cancer incidence ([100, 101](#)).

### **Colorectal cancer**

A pooled analysis of 13 [prospective cohort studies](#), which followed a total of 725,134 individuals for a 7 to 20-year period, revealed a modest, inverse association between dietary and total (from food and [supplements](#)) folate intake and [colon cancer](#) risk. Specifically, a 2% decrease in colon cancer risk was estimated for every 100 µg/day increase in total folate intake ([102](#)). A large US [prospective study](#), which followed 525,488 subjects, ages 50 to 71 years between 1995 and 2006, correlated dietary folate, supplemental folic acid, and total folate intakes with a decreased colorectal cancer (CRC) risk ([103](#)). However, when stratified by gender, there was no association between dietary folate intake and CRC risk in women ([103, 104](#)). A lack of association between CRC risk and dietary, supplemental, and total folate intakes was also reported in another prospective study that followed more than 90,000 US postmenopausal women during an 11-year period encompassing pre- and post-[fortification](#) periods ([105](#)). These data suggest the possible influence of gender over CRC risk modification by folate. In the latter study, a significant but transient risk elevation was also observed during the post-fortification era; however, some have asserted that this is unlikely to be caused by increased folate intake due to mandatory fortification ([106](#)). Finally, a [meta-analysis](#) of 18 [case-control studies](#) found a slight reduction in CRC risk with folate from food ([107](#)). However, it is important to note that the case-control studies were highly [heterogeneous](#), and that the authors stated that dietary fiber, vitamins, and alcohol intake could have confounded their results. Moreover, the lower limit of the highest quantile of folate intake was highly variable, ranging from 270 to 1,367 µg/day ([107](#)).

While most [epidemiological](#) research shows a protective effect of folate against colorectal cancer development, it has been suggested that high doses of supplemental folic acid may actually accelerate tumor growth in cancer patients ([108](#)). Whereas higher folate status within the normal dietary range is widely considered to be protective against cancer, some investigators remain concerned that exposure to excessively high folic acid intakes may increase the growth of pre-existing neoplasms ([108](#)). Several [clinical trials](#) addressed the effect of folic acid supplementation in patients with a history of [colorectal adenoma](#), with trials finding a risk reduction or no effect of supplemental folic acid ([109-112](#)). A recent meta-analysis of three large [randomized controlled trials](#) in high-risk subjects did not demonstrate any increase in colorectal adenoma recurrence in subjects supplemented with 500 or 1,000 µg/day of folic acid for 24 to 42 months when compared with [placebo](#) treatment ([113](#)).

As suggested earlier, the *MTHFR* 677T/T genotype might prevent uracil misincorporation and protect [DNA](#) integrity and stability under low-folate conditions. A meta-analysis of 62 case-control and two cohort studies revealed that while the T/T variant reduces CRC risk by 12% compared to both C/T and C/C genotypes, the risk was decreased by 30% with high (348-1,583 µg/day) versus low total folate intakes (264-450 µg/day), irrespective of the genotype ([114](#)). A common [polymorphism](#) (c.2756A>G) in the *MTR* gene, which codes for methionine synthase, was also examined in relation with the risk of colorectal adenoma and cancer. Methionine synthase [catalyzes](#) the simultaneous conversion of [homocysteine](#) and 5-methylene THF into methionine and TFH, respectively. The recent meta-analysis of 27 case-control studies showed no association between *MTR* variant and cancer risk ([115](#)).

Although alcohol consumption interferes with the absorption and metabolism of folate (16), one case-control and five prospective cohort studies have reported either reduction in CRC risk among nondrinkers compared to drinkers or a lack of association (107). However, in a large prospective study that followed more than 28,000 male health professionals for 22 years, intake of more than two alcoholic drinks (>30 grams of alcohol) per day augmented CRC risk by 42% during the pre-fortification period. CRC risk was not increased during the post-fortification period, suggesting that it is the combination of high alcohol and low folate intake that might increase CRC risk. Yet, another prospective study that followed more than 69,000 female nurses for 28 years did not report a significant increase in CRC risk with alcohol intake before and after the mandatory folic acid fortification (116). In some studies, individuals who are [homozygous](#) for the c.677C>T *MTHFR* polymorphism (T/T) have been found to be at decreased risk for colon cancer when folate intake is adequate. However, when folate intake is low and/or alcohol intake is high, individuals with the (T/T) genotype have been found to be at increased risk of colorectal cancer (117, 118).

### ***Breast cancer***

Several [prospective cohort](#) and [case-control studies](#) investigating whether folate intake affects breast [cancer risk](#) have reported mixed results (119). A [meta-analysis](#) of 15 prospective studies and one [nested case-control study](#) found no relationship with dietary folate intake (120). Moderate alcohol intake has been associated with increased [risk](#) of breast cancer in women (121). The results of three prospective studies suggested that increased folate intake may reduce the risk of breast cancer in women who regularly consume alcohol (122-124). Thus, high folate intake might be associated with a risk reduction only in women whose breast cancer risk is raised by alcohol consumption. A very large prospective study in more than 88,000 nurses reported that folic acid intake was not associated with breast cancer in women who consumed less than one alcoholic drink per day. However, in women consuming at least one alcoholic drink per day, folic acid intake of at least 600 µg daily resulted in about half the risk of breast cancer compared with women who consumed less than 300 µg of folic acid daily (124). Nevertheless, whether and how alcohol consumption increases breast cancer risk is still subject to discussion (125, 126). Finally, recent meta-analyses evaluating the influence of [polymorphisms](#) in [one-carbon metabolism](#) on cancer risk found that specific variants in the [gene](#) encoding thymidylate synthase increased the risk of breast cancer in certain ethnic populations (127, 128).

### ***Childhood cancers***

The incidence of Wilms' tumors (kidney [cancer](#)) and certain types of brain cancers (neuroblastoma, ganglioneuroblastoma, and ependymoma) in children has decreased since the mandatory [fortification](#) of the US grain supply in 1998 (129). However, incidence rates were unchanged between the pre- and post-fortification periods for leukemia—a predominant childhood malignancy. Despite earlier studies linking maternal folic acid supplementation during pregnancy with the reduced [risk](#) of childhood leukemia, more recent investigations have found little evidence to support a preventive effect of folic acid (130). Several [meta-analyses](#) have also found little to no protective effect with *MTHFR* [polymorphisms](#); however, the most recent meta-analysis of 22 [case-control studies](#) found a reduction in the risk of acute lymphoblastic [leukemia](#) (ALL) with the c.677C>T variant in Caucasians and Asians (131).

### **Alzheimer's disease and cognitive impairment**

[Alzheimer's disease](#) (AD) is the most common form of [dementia](#), affecting more than 5 million individuals over 65 years old in the US (132). [β-amyloid plaque](#) deposition, Tau protein-forming tangles, and increased cell death in the brain of AD patients have been associated with [cognitive](#) decline and memory loss. One study associated increased consumption of fruit and vegetables, which are abundant sources of folate, with a reduced [risk](#) of developing dementia and AD in women (133). Through its role in [nucleic acid synthesis](#) and methyl donor provision for [methylation](#) reactions, folate is critical for normal brain development and function, not only during pregnancy and after birth, but also later in life (134). In one [cross-sectional study](#) of elderly women, AD patients had significantly higher [homocysteine](#) and lower red blood cell folate concentrations compared to healthy individuals. However, there was no difference in the level of [serum](#) folate between

groups, suggesting that long-term folate status, rather than recent folate intake, may be associated with the risk of AD (135).

Several investigators have described associations between increased homocysteine concentrations and cognitive impairment in the elderly (136), but [prospective cohort studies](#) have not found higher folate intakes to be associated with improved [cognition](#) (137, 138). Higher homocysteine concentrations were found in individuals suffering from dementia, including AD and [vascular dementia](#), compared to healthy subjects (139, 140). Although deficiencies in folate, vitamin B<sub>12</sub>, and vitamin B<sub>6</sub> could increase homocysteine concentrations, a reduction in [vitamin](#) concentrations in the serum of AD patients compared to healthy individuals could not be attributed to decreased vitamin intakes (141). It is not presently clear whether serum homocysteine is a risk factor for developing dementia or simply associated with the cognitive decline. In the last decade, a number of [clinical trials](#) have tested the use of B-vitamins to lower homocysteine and prevent or delay cognitive decline. A meta-analysis of nine randomized, placebo-controlled trials of folic acid supplementation (0.2 to 15 mg/day for a median duration of six months) in healthy individuals over 45 years of age failed to find a short-term effect on cognitive functions, including memory, speed, language, and [executive functions](#) (142). More recently, a [meta-analysis](#) of 19 [randomized, placebo-controlled](#) trials of B-vitamin supplementation found no difference in cognitive parameters between the treatment and placebo groups, despite the treatment effectively lowering homocysteine concentrations (143). Inconsistent findings across trials may be due to differences in design and methodology (reviewed in 144).

Nevertheless, a two-year randomized, placebo-controlled trial in 168 elderly subjects with mild cognitive impairment recently described the benefits of a daily regimen of 800 µg of folic acid, 500 µg of vitamin B<sub>12</sub>, and 20 mg of vitamin B<sub>6</sub> (145, 146). [Atrophy](#) of specific brain regions affected by AD was observed in individuals of both groups, and this atrophy correlated with cognitive decline; however, the B-vitamin treatment group experienced a smaller loss of [gray matter](#) compared to the placebo group (0.5% vs. 3.7%). A greater benefit was seen in subjects with higher baseline homocysteine concentrations, suggesting the importance of lowering circulating homocysteine in prevention of cognitive decline and dementia. Although encouraging, the effect of B-vitamin supplementation needs to be further studied in larger trials that evaluate long-term outcomes, such as the incidence of AD.

## Disease Treatment

### Metabolic diseases

Folinic acid (see [Figure 1](#) above), a tetrahydrofolic acid derivative, is used in the clinical management of rare inborn errors that affect folate transport or [metabolism](#) (reviewed in 147). Such conditions are of [autosomal recessive](#) inheritance, meaning only individuals receiving two copies of the mutated [gene](#) (one from each parent) develop the disease.

#### *Hereditary folate malabsorption*

Hereditary folate [malabsorption](#) is caused by [mutations](#) in the *SLC46A1* [gene](#) coding for the folate transporter PCFT and typically affects [gastrointestinal](#) folate absorption and folate transport into the brain (148). Patients present with low to undetectable concentrations of folate in [serum](#) and [cerebrospinal fluid](#), pancytopenia (low number of all blood cells), impaired immune responses that increase susceptibility to infections, and a general failure to thrive (149). [Neurologic](#) symptoms, including [seizures](#), have also been observed (150). Clinical improvements have been recorded following [parenteral](#) provision of folinic acid (151).

#### *Cerebral Folate Deficiency (CFD) syndrome*

CFD is characterized by low levels of folate [coenzymes](#) in [cerebrospinal fluid](#) despite normal concentrations of folate in blood. Folate transport across the blood-brain barrier is compromised in CFD and has been linked

either to the presence of [antibodies](#) blocking the folate receptor FR $\alpha$  or to [mutations](#) in the *FOLR1* [gene](#) encoding FR $\alpha$  ([152, 153](#)). [Neurologic](#) abnormalities, along with visual and hearing impairments, have been described in children with CFD; autism spectrum disorder (ASD) is present in some cases. Folinic acid (also known as leucovorin) can enter the brain and normalize the level of folate coenzymes and has been shown to normalize folate concentrations and improve various social interactions in CFD, including mood, behavior, and verbal communication in children with ASD ([152, 154, 155](#)).

### *Dihydrofolate reductase (DHFR) deficiency*

DHFR is the NADPH-dependent [enzyme](#) that [catalyzes](#) the reduction of dihydrofolic acid (DHF) to tetrahydrofolic acid (THF). DHFR is also required to convert folic acid to DHF. DHFR deficiency is characterized by [megaloblastic anemia](#) and [cerebral](#) folate deficiency causing intractable [seizures](#) and mental deficits. Although folinic acid treatment can alleviate the symptoms of DHFR deficiency, early diagnosis is essential to prevent irreversible brain damage and improve clinical outcomes ([156, 157](#)).

## Sources

### Food sources

Green leafy vegetables (foliage) are rich sources of folate and provide the basis for its name. Citrus fruit juices, [legumes](#), and [fortified](#) foods are also excellent sources of folate ([1](#)); the folate content of fortified cereal varies greatly. A number of folate-rich foods are listed in **Table 2**, along with their folate content in micrograms ( $\mu\text{g}$ ). For more information on the nutrient content of specific foods, search the [USDA food composition database](#).

Table 2. Some Food Sources of Folate and Folic Acid

Food	Serving	Folate ( $\mu\text{g}$ DFEs)
Lentils (mature seeds, cooked, boiled)	$\frac{1}{2}$ cup	179
Garbanzo beans (chickpeas, cooked, boiled)	$\frac{1}{2}$ cup	141
Asparagus (cooked, boiled)	$\frac{1}{2}$ cup (~6 spears)	134
Spinach (cooked, boiled)	$\frac{1}{2}$ cup	131
Lima beans (large, mature seeds, cooked, boiled)	$\frac{1}{2}$ cup	78
Orange juice (raw)	6 fl. oz.	56

\*To help prevent neural tube defects, the US FDA required the addition of 1.4 milligrams (mg) of folic acid per kilogram (kg) of grain to be added to refined grain products, which are already enriched with niacin, thiamin, riboflavin, and iron, as of January 1, 1998. The addition of nutrients to food in order to prevent a nutritional deficiency or restore nutrients lost in processing is known as [fortification](#). The FDA initially estimated that this level of fortification would increase dietary intake by an average of 100  $\mu\text{g}$  folic acid/day ([26](#)). However, further evaluations based on [observational studies](#) suggested increases twice that predicted by the FDA ([35](#)). The prevalence of low folate concentrations in both [serum](#) and red blood cells is currently below 1% in the US population, compared to 24% and 3.5%, respectively, before the fortification period ([158](#)).

Spaghetti (enriched, cooked)	1 cup	167*
White rice (enriched, cooked)	1 cup	153*
Bread (enriched)	1 slice	84*

\*To help prevent neural tube defects, the US FDA required the addition of 1.4 milligrams (mg) of folic acid per kilogram (kg) of grain to be added to refined grain products, which are already enriched with niacin, thiamin, riboflavin, and iron, as of January 1, 1998. The addition of nutrients to food in order to prevent a nutritional deficiency or restore nutrients lost in processing is known as [fortification](#). The FDA initially estimated that this level of fortification would increase dietary intake by an average of 100 µg folic acid/day (26). However, further evaluations based on [observational studies](#) suggested increases twice that predicted by the FDA (35). The prevalence of low folate concentrations in both [serum](#) and red blood cells is currently below 1% in the US population, compared to 24% and 3.5%, respectively, before the fortification period (158).

## Supplements

The principal form of [supplementary](#) folate is folic acid. It is available in single-ingredient and combination products, such as B-complex vitamins and multivitamins. Doses of 1 mg or greater require a prescription (159). Additionally, folinic acid, a tetrahydrofolic acid derivative, is used to manage certain metabolic diseases (see [Disease Treatment](#)). Further, the US FDA has approved the supplementation of folate in oral contraceptives. The addition of levomefolate calcium (the calcium salt of MeTHF; 451 µg/tablet) to oral contraceptives is intended to raise folate status in women of childbearing age (160). According to a US national survey, only 24% of non-pregnant women aged 15-44 years are meeting the current recommendation of 400 µg/day of folic acid (161).

## Safety

### Toxicity

No adverse effects have been associated with the consumption of excess folate from food. Concerns regarding safety are limited to synthetic folic acid intake. Deficiency of vitamin B<sub>12</sub>, though often undiagnosed, may affect a significant number of people, especially older adults (see the article on [Vitamin B<sub>12</sub>](#)). One symptom of vitamin B<sub>12</sub> deficiency is [megaloblastic anemia](#), which is indistinguishable from that associated with folate deficiency (see [Deficiency](#)). Large doses of folic acid given to an individual with an undiagnosed vitamin B<sub>12</sub> deficiency could correct megaloblastic anemia without correcting the underlying vitamin B<sub>12</sub> deficiency, leaving the individual at [risk](#) of developing irreversible [neurologic](#) damage. Such cases of neurologic progression in vitamin B<sub>12</sub> deficiency have been mostly seen at folic acid doses of 5,000 µg (5 mg) and above. In order to be very sure of preventing irreversible neurological damage in vitamin B<sub>12</sub>-deficient individuals, the Food and Nutrition Board of the US Institute of Medicine advises that all adults limit their intake of folic acid ([supplements](#) and [fortification](#)) to 1,000 µg (1 mg) daily (**Table 3**). The Board also noted that vitamin B<sub>12</sub> deficiency is very rare in women in their childbearing years, making the consumption of folic acid at or above 1,000 µg/day unlikely to cause problems (1); however, there are limited data on the effects of large doses.

Table 3. Tolerable Upper Intake Level (UL) for Folic Acid

Age Group	UL (µg/day)
*Source of intake should be from food and formula only.	

Infants 0-12 months	Not possible to establish*
Children 1-3 years	300
Children 4-8 years	400
Children 9-13 years	600
Adolescents 14-18 years	800
Adults 19 years and older	1,000
*Source of intake should be from food and formula only.	

The saturation of DHFR metabolic capacity by oral doses of folic acid has been associated with the appearance of unmetabolized folic acid in blood (162). [Hematologic](#) abnormalities and poorer [cognition](#) have been associated with the presence of unmetabolized folic acid in vitamin B<sub>12</sub>-deficient older adults (≥60 years) (163, 164). A small study conducted in postmenopausal women also raised concerns about the effect of exposure to unmetabolized folic acid on immune function (165). In a small, [randomized, open-label trial](#) in 38 women of reproductive age receiving 30 weeks of daily multivitamin supplements, daily supplementation with either 1.1 mg or 5 mg of folic acid resulted in the transient appearance of unmetabolized folic acid in blood over the first 12 weeks of supplementation (166). However, unmetabolized folic acid concentrations returned to baseline levels at the end of the study, suggesting that adaptive mechanisms eventually converted folic acid to reduced forms of folate. Nonetheless, the use of supplemental levomefolate (5-methyl THF) may provide an alternative to prevent the potential negative effects of unconverted folic acid in older adults.

### Drug interactions

When nonsteroidal anti-[inflammatory](#) drugs (NSAIDs), such as aspirin or ibuprofen, are taken in very large therapeutic dosages (i.e., to treat severe arthritis), they may interfere with folate [metabolism](#). In contrast, routine use of NSAIDs has not been found to adversely affect folate [status](#). The [anticonvulsant](#), phenytoin, has been shown to inhibit the [intestinal](#) absorption of folate, and several studies have associated decreased folate status with long-term use of the anticonvulsants, phenytoin, phenobarbital, and primidone (167). However, few studies controlled for differences in dietary folate intake between anticonvulsant users and nonusers. Also, taking folic acid at the same time as the [cholesterol](#)-lowering agents, cholestyramine and colestipol, may decrease the absorption of folic acid (159). Methotrexate is a folate [antagonist](#) used to treat a number of diseases, including [cancer](#), [rheumatoid arthritis](#), and [psoriasis](#). Some of the side effects of methotrexate are similar to those of severe folate deficiency, and supplementation with folic or folinic acid is used to reduce antifolate toxicity. Other antifolate molecules currently used in cancer therapy include aminopterin, pemetrexed, pralatrexate, and raltitrexed (96). Further, a number of other medications have been shown to have antifolate activity, including trimethoprim (an antibiotic), pyrimethamine (an antimalarial), triamterene (a blood pressure medication), and sulfasalazine (a treatment for [ulcerative colitis](#)). Early studies of oral contraceptives (birth control pills) containing high doses of [estrogen](#) indicated adverse effects on folate status; however, this finding has not been supported in more recent studies that used low-dose oral contraceptives and controlled for dietary folate (168).

### Linus Pauling Institute Recommendation

The available scientific evidence shows that adequate folate intake prevents [neural tube defects](#) and other poor outcomes of pregnancy; is helpful in lowering the [risk](#) of some forms of [cancer](#), especially in genetically susceptible individuals; and may lower the risk of [cardiovascular disease](#). The Linus Pauling Institute recommends that adults take a daily multivitamin/mineral supplement, which typically contains 400 µg of folic acid, the Daily Value (DV). Even with a larger than average intake of folic acid from [fortified](#) food, it is

unlikely that an individual's daily folic acid intake would regularly exceed the tolerable upper intake level of 1,000 µg/day established by the Institute of Medicine (see [Safety](#)).

### Older adults (>50 years)

The recommendation for 400 µg/day of [supplemental](#) folic acid as part of a daily multivitamin/mineral supplement, in addition to a folate-rich diet, is especially important for older adults because blood [homocysteine](#) concentrations tend to increase with age (see [Disease Prevention](#)).

---

## Authors and Reviewers

Originally written in 2000 by:

Jane Higdon, Ph.D.

Linus Pauling Institute

Oregon State University

Updated in April 2002 by:

Jane Higdon, Ph.D.

Linus Pauling Institute

Oregon State University

Updated in September 2007 by:

Victoria J. Drake, Ph.D.

Linus Pauling Institute

Oregon State University

Updated in June 2014 by:

Barbara Delage, Ph.D.

Linus Pauling Institute

Oregon State University

Reviewed in December 2014 by:

Helene McNulty, Ph.D., R.D.

Professor of Human Nutrition and Dietetics

Northern Ireland Centre for Food and Health (NICHE)

University of Ulster

Coleraine, United Kingdom

The 2014 update of this article was underwritten, in part, by a grant from [Bayer Consumer Care AG](#), Basel, Switzerland.

Copyright 2000-2018 Linus Pauling Institute

---

## References

1. Food and Nutrition Board, Institute of Medicine. Folate. Dietary Reference Intakes: Thiamin, Riboflavin, Niacin, Vitamin B<sub>6</sub>, Folate, Vitamin B<sub>12</sub>, Pantothenic Acid, Biotin, and Choline. Washington, D.C.: National Academy Press; 1998:196-305. ([National Academy Press](#))
2. Choi SW, Mason JB. Folate and carcinogenesis: an integrated scheme. J Nutr. 2000;130(2):129-132. ([PubMed](#))

3. Bailey LB, Gregory JF, 3rd. Folate metabolism and requirements. *J Nutr.* 1999;129(4):779-782. ([PubMed](#))
4. Gerhard GT, Duell PB. Homocysteine and atherosclerosis. *Curr Opin Lipidol.* 1999;10(5):417-428. ([PubMed](#))
5. Jacques PF, Bostom AG, Wilson PW, Rich S, Rosenberg IH, Selhub J. Determinants of plasma total homocysteine concentration in the Framingham Offspring cohort. *Am J Clin Nutr.* 2001;73(3):613-621. ([PubMed](#))
6. Jacques PF, Kalmbach R, Bagley PJ, et al. The relationship between riboflavin and plasma total homocysteine in the Framingham Offspring cohort is influenced by folate status and the C677T transition in the methylenetetrahydrofolate reductase gene. *J Nutr.* 2002;132(2):283-288. ([PubMed](#))
7. McNulty H, Dowey le RC, Strain JJ, et al. Riboflavin lowers homocysteine in individuals homozygous for the MTHFR 677C->T polymorphism. *Circulation.* 2006;113(1):74-80. ([PubMed](#))
8. Verlinde PH, Oey I, Hendrickx ME, Van Loey AM, Temme EH. L-ascorbic acid improves the serum folate response to an oral dose of [6S]-5-methyltetrahydrofolic acid in healthy men. *Eur J Clin Nutr.* 2008;62(10):1224-1230. ([PubMed](#))
9. Lucock M, Yates Z, Boyd L, et al. Vitamin C-related nutrient-nutrient and nutrient-gene interactions that modify folate status. *Eur J Nutr.* 2013;52(2):569-582. ([PubMed](#))
10. Desmoulin SK, Hou Z, Gangjee A, Matherly LH. The human proton-coupled folate transporter: Biology and therapeutic applications to cancer. *Cancer Biol Ther.* 2012;13(14):1355-1373. ([PubMed](#))
11. Solanky N, Requena Jimenez A, D'Souza SW, Sibley CP, Glazier JD. Expression of folate transporters in human placenta and implications for homocysteine metabolism. *Placenta.* 2010;31(2):134-143. ([PubMed](#))
12. Halsted CH, Villanueva JA, Devlin AM, Chandler CJ. Metabolic interactions of alcohol and folate. *J Nutr.* 2002;132(8 Suppl):2367S-2372S. ([PubMed](#))
13. Pfeiffer CM, Sternberg MR, Schleicher RL, Rybak ME. Dietary supplement use and smoking are important correlates of biomarkers of water-soluble vitamin status after adjusting for sociodemographic and lifestyle variables in a representative sample of US adults. *J Nutr.* 2013;143(6):957S-965S. ([PubMed](#))
14. Stark KD, Pawlosky RJ, Sokol RJ, Hannigan JH, Salem N, Jr. Maternal smoking is associated with decreased 5-methyltetrahydrofolate in cord plasma. *Am J Clin Nutr.* 2007;85(3):796-802. ([PubMed](#))
15. Hutson JR, Stade B, Lehotay DC, Collier CP, Kapur BM. Folic acid transport to the human fetus is decreased in pregnancies with chronic alcohol exposure. *PLoS One.* 2012;7(5):e38057. ([PubMed](#))
16. Herbert V. Folic acid. In: Shils M, Olson JA, Shike M, Ross AC, eds. *Modern Nutrition in Health and Disease.* 9th ed. Baltimore: Lippincott Williams & Wilkins; 1999:433-446.
17. Stabler SP. Clinical folate deficiency. In: Bailey LB, ed. *Folate in Health and Disease.* 2nd edition ed. Boca Raton, FL: CRC press, Taylor & Francis Group; 2010:409-428.
18. Bailey LB. Dietary reference intakes for folate: the debut of dietary folate equivalents. *Nutr Rev.* 1998;56(10):294-299. ([PubMed](#))
19. Bailey LB, Gregory JF, 3rd. Polymorphisms of methylenetetrahydrofolate reductase and other enzymes: metabolic significance, risks and impact on folate requirement. *J Nutr.* 1999;129(5):919-922. ([PubMed](#))
20. Wilcken B, Bamforth F, Li Z, et al. Geographical and ethnic variation of the 677C>T allele of 5,10 methylenetetrahydrofolate reductase (MTHFR): findings from over 7000 newborns from 16 areas world wide.

J Med Genet. 2003;40(8):619-625. ([PubMed](#))

21. Guenther BD, Sheppard CA, Tran P, Rozen R, Matthews RG, Ludwig ML. The structure and properties of methylenetetrahydrofolate reductase from *Escherichia coli* suggest how folate ameliorates human hyperhomocysteinemia. *Nat Struct Biol.* 1999;6(4):359-365. ([PubMed](#))

22. Molloy AM, Daly S, Mills JL, et al. Thermolabile variant of 5,10-methylenetetrahydrofolate reductase associated with low red-cell folates: implications for folate intake recommendations. *Lancet.* 1997;349(9065):1591-1593. ([PubMed](#))

23. Rozen R. Genetic predisposition to hyperhomocysteinemia: deficiency of methylenetetrahydrofolate reductase (MTHFR). *Thromb Haemost.* 1997;78(1):523-526. ([PubMed](#))

24. Kauwell GP, Wilsky CE, Cerda JJ, et al. Methylenetetrahydrofolate reductase mutation (677C-->T) negatively influences plasma homocysteine response to marginal folate intake in elderly women. *Metabolism.* 2000;49(11):1440-1443. ([PubMed](#))

25. Shane B. Folic acid, vitamin B-12, and vitamin B-6. In: Stipanuk M, ed. *Biochemical and Physiological Aspects of Human Nutrition.* Philadelphia: W.B. Saunders Co.; 2000:483-518.

26. Eskes TK. Open or closed? A world of difference: a history of homocysteine research. *Nutr Rev.* 1998;56(8):236-244. ([PubMed](#))

27. Czeizel AE, Dudas I, Vereczkey A, Banhidy F. Folate deficiency and folic acid supplementation: the prevention of neural-tube defects and congenital heart defects. *Nutrients.* 2013;5(11):4760-4775. ([PubMed](#))

28. MRC Vitamin Study Research Group. Prevention of neural tube defects: results of the Medical Research Council Vitamin Study. *Lancet.* 1991;338(8760):131-137. ([PubMed](#))

29. Czeizel AE, Dudas I. Prevention of the first occurrence of neural-tube defects by periconceptional vitamin supplementation. *N Engl J Med.* 1992;327(26):1832-1835. ([PubMed](#))

30. Talaulikar VS, Arulkumaran S. Folic acid in obstetric practice: a review. *Obstet Gynecol Surv.* 2011;66(4):240-247. ([PubMed](#))

31. American College of Obstetricians and Gynecologists (ACOG). Neural tube defects. Washington, DC. 2003. Available at: <http://www.guideline.gov/content.aspx?id=3994>. Accessed 12/19/14.

32. McNulty B, Pentieva K, Marshall B, et al. Women's compliance with current folic acid recommendations and achievement of optimal vitamin status for preventing neural tube defects. *Hum Reprod.* 2011;26(6):1530-1536. ([PubMed](#))

33. Nilsen RM, Vollset SE, Gjessing HK, et al. Patterns and predictors of folic acid supplement use among pregnant women: the Norwegian Mother and Child Cohort Study. *Am J Clin Nutr.* 2006;84(5):1134-1141. ([PubMed](#))

34. Ray JG, Singh G, Burrows RF. Evidence for suboptimal use of periconceptional folic acid supplements globally. *BJOG.* 2004;111(5):399-408. ([PubMed](#))

35. Quinlivan EP, Gregory JF, 3rd. Effect of food fortification on folic acid intake in the United States. *Am J Clin Nutr.* 2003;77(1):221-225. ([PubMed](#))

36. National Birth Defects Prevention Network. Neural Tube Defect Ascertainment Project. Available at: [http://www.nbdpn.org/ntd\\_folic\\_acid\\_information.php](http://www.nbdpn.org/ntd_folic_acid_information.php). Accessed 12/16/14.

37. Copp AJ, Stanier P, Greene ND. Neural tube defects: recent advances, unsolved questions, and controversies. *Lancet Neurol.* 2013;12(8):799-810. ([PubMed](#))
38. Yan L, Zhao L, Long Y, et al. Association of the maternal MTHFR C677T polymorphism with susceptibility to neural tube defects in offsprings: evidence from 25 case-control studies. *PLoS One.* 2012;7(10):e41689. ([PubMed](#))
39. De Marco P, Calevo MG, Moroni A, et al. Study of MTHFR and MS polymorphisms as risk factors for NTD in the Italian population. *J Hum Genet.* 2002;47(6):319-324. ([PubMed](#))
40. van der Put NM, Gabreels F, Stevens EM, et al. A second common mutation in the methylenetetrahydrofolate reductase gene: an additional risk factor for neural-tube defects? *Am J Hum Genet.* 1998;62(5):1044-1051. ([PubMed](#))
41. De Marco P, Calevo MG, Moroni A, et al. Reduced folate carrier polymorphism (80A-->G) and neural tube defects. *Eur J Hum Genet.* 2003;11(3):245-252. ([PubMed](#))
42. O'Leary VB, Mills JL, Parle-McDermott A, et al. Screening for new MTHFR polymorphisms and NTD risk. *Am J Med Genet A.* 2005;138A(2):99-106. ([PubMed](#))
43. Christensen B, Arbour L, Tran P, et al. Genetic polymorphisms in methylenetetrahydrofolate reductase and methionine synthase, folate levels in red blood cells, and risk of neural tube defects. *Am J Med Genet.* 1999;84(2):151-157. ([PubMed](#))
44. Relton CL, Wilding CS, Pearce MS, et al. Gene-gene interaction in folate-related genes and risk of neural tube defects in a UK population. *J Med Genet.* 2004;41(4):256-260. ([PubMed](#))
45. Brody LC, Conley M, Cox C, et al. A polymorphism, R653Q, in the trifunctional enzyme methylenetetrahydrofolate dehydrogenase/methenyltetrahydrofolate cyclohydrolase/formyltetrahydrofolate synthetase is a maternal genetic risk factor for neural tube defects: report of the Birth Defects Research Group. *Am J Hum Genet.* 2002;71(5):1207-1215. ([PubMed](#))
46. van der Put NM, van den Heuvel LP, Steegers-Theunissen RP, et al. Decreased methylene tetrahydrofolate reductase activity due to the 677C-->T mutation in families with spina bifida offspring. *J Mol Med (Berl).* 1996;74(11):691-694. ([PubMed](#))
47. Wilson A, Platt R, Wu Q, et al. A common variant in methionine synthase reductase combined with low cobalamin (vitamin B12) increases risk for spina bifida. *Mol Genet Metab.* 1999;67(4):317-323. ([PubMed](#))
48. Gilboa SM, Salemi JL, Nembhard WN, Fixler DE, Correa A. Mortality resulting from congenital heart disease among children and adults in the United States, 1999 to 2006. *Circulation.* 2010;122(22):2254-2263. ([PubMed](#))
49. van Beynum IM, Kapusta L, Bakker MK, den Heijer M, Blom HJ, de Walle HE. Protective effect of periconceptional folic acid supplements on the risk of congenital heart defects: a registry-based case-control study in the northern Netherlands. *Eur Heart J.* 2010;31(4):464-471. ([PubMed](#))
50. Yin M, Dong L, Zheng J, Zhang H, Liu J, Xu Z. Meta analysis of the association between MTHFR C677T polymorphism and the risk of congenital heart defects. *Ann Hum Genet.* 2012;76(1):9-16. ([PubMed](#))
51. Wang W, Wang Y, Gong F, Zhu W, Fu S. MTHFR C677T polymorphism and risk of congenital heart defects: evidence from 29 case-control and TDT studies. *PLoS One.* 2013;8(3):e58041. ([PubMed](#))
52. Badovinac RL, Werler MM, Williams PL, Kelsey KT, Hayes C. Folic acid-containing supplement consumption during pregnancy and risk for oral clefts: a meta-analysis. *Birth Defects Res A Clin Mol Teratol.* 2007;79(1):8-15. ([PubMed](#))

53. Wilcox AJ, Lie RT, Solvoll K, et al. Folic acid supplements and risk of facial clefts: national population based case-control study. *BMJ*. 2007;334(7591):464. ([PubMed](#))
54. Boyles AL, Wilcox AJ, Taylor JA, et al. Folate and one-carbon metabolism gene polymorphisms and their associations with oral facial clefts. *Am J Med Genet A*. 2008;146A(4):440-449. ([PubMed](#))
55. Boyles AL, Wilcox AJ, Taylor JA, et al. Oral facial clefts and gene polymorphisms in metabolism of folate/one-carbon and vitamin A: a pathway-wide association study. *Genet Epidemiol*. 2009;33(3):247-255. ([PubMed](#))
56. Luo YL, Cheng YL, Ye P, Wang W, Gao XH, Chen Q. Association between MTHFR polymorphisms and orofacial clefts risk: a meta-analysis. *Birth Defects Res A Clin Mol Teratol*. 2012;94(4):237-244. ([PubMed](#))
57. Wilcox AJ. On the importance--and the unimportance--of birthweight. *Int J Epidemiol*. 2001;30(6):1233-1241. ([PubMed](#))
58. Fekete K, Berti C, Trovato M, et al. Effect of folate intake on health outcomes in pregnancy: a systematic review and meta-analysis on birth weight, placental weight and length of gestation. *Nutr J*. 2012;11:75. ([PubMed](#))
59. Baker PN, Wheeler SJ, Sanders TA, et al. A prospective study of micronutrient status in adolescent pregnancy. *Am J Clin Nutr*. 2009;89(4):1114-1124. ([PubMed](#))
60. Lee HA, Park EA, Cho SJ, et al. Mendelian randomization analysis of the effect of maternal homocysteine during pregnancy, as represented by maternal MTHFR C677T genotype, on birth weight. *J Epidemiol*. 2013;23(5):371-375. ([PubMed](#))
61. Scholl TO, Johnson WG. Folic acid: influence on the outcome of pregnancy. *Am J Clin Nutr*. 2000;71(5 Suppl):1295S-1303S. ([PubMed](#))
62. Vollset SE, Refsum H, Irgens LM, et al. Plasma total homocysteine, pregnancy complications, and adverse pregnancy outcomes: the Hordaland Homocysteine study. *Am J Clin Nutr*. 2000;71(4):962-968. ([PubMed](#))
63. Wang XM, Wu HY, Qiu XJ. Methylenetetrahydrofolate reductase (MTHFR) gene C677T polymorphism and risk of preeclampsia: an updated meta-analysis based on 51 studies. *Arch Med Res*. 2013;44(3):159-168. ([PubMed](#))
64. Wen SW, Champagne J, Rennicks White R, et al. Effect of folic acid supplementation in pregnancy on preeclampsia: the folic acid clinical trial study. *J Pregnancy*. 2013;2013:294312. ([PubMed](#))
65. Lassi ZS, Salam RA, Haider BA, Bhutta ZA. Folic acid supplementation during pregnancy for maternal health and pregnancy outcomes. *Cochrane Database Syst Rev*. 2013;3:CD006896. ([PubMed](#))
66. Schmidt RJ, Tancredi DJ, Ozonoff S, et al. Maternal periconceptional folic acid intake and risk of autism spectrum disorders and developmental delay in the CHARGE (CHILDhood Autism Risks from Genetics and Environment) case-control study. *Am J Clin Nutr*. 2012;96(1):80-89. ([PubMed](#))
67. Crider KS, Cordero AM, Qi YP, Mulinare J, Dowling NF, Berry RJ. Prenatal folic acid and risk of asthma in children: a systematic review and meta-analysis. *Am J Clin Nutr*. 2013;98(5):1272-1281. ([PubMed](#))
68. Brown SB, Reeves KW, Bertone-Johnson ER. Maternal folate exposure in pregnancy and childhood asthma and allergy: a systematic review. *Nutr Rev*. 2014;72(1):55-64. ([PubMed](#))
69. Ding R, Lin S, Chen D. The association of cystathionine  $\beta$  synthase (CBS) T833C polymorphism and the risk of stroke: a meta-analysis. *J Neurol Sci*. 2012;312(1-2):26-30. ([PubMed](#))

70. Seshadri N, Robinson K. Homocysteine, B vitamins, and coronary artery disease. *Med Clin North Am.* 2000;84(1):215-237. ([PubMed](#))
71. Wald DS, Law M, Morris JK. Homocysteine and cardiovascular disease: evidence on causality from a meta-analysis. *BMJ.* 2002;325(7374):1202. ([PubMed](#))
72. Homocysteine Studies Collaboration. Homocysteine and risk of ischemic heart disease and stroke: a meta-analysis. *JAMA.* 2002;288(16):2015-2022. ([PubMed](#))
73. Clarke R, Halsey J, Bennett D, Lewington S. Homocysteine and vascular disease: review of published results of the homocysteine-lowering trials. *J Inherit Metab Dis.* 2011;34(1):83-91. ([PubMed](#))
74. Huang T, Chen Y, Yang B, Yang J, Wahlqvist ML, Li D. Meta-analysis of B vitamin supplementation on plasma homocysteine, cardiovascular and all-cause mortality. *Clin Nutr.* 2012;31(4):448-454. ([PubMed](#))
75. Voutilainen S, Rissanen TH, Virtanen J, Lakka TA, Salonen JT. Low dietary folate intake is associated with an excess incidence of acute coronary events: The Kuopio Ischemic Heart Disease Risk Factor Study. *Circulation.* 2001;103(22):2674-2680. ([PubMed](#))
76. Brattstrom L. Vitamins as homocysteine-lowering agents. *J Nutr.* 1996;126(4 Suppl):1276S-1280S. ([PubMed](#))
77. Rader JI. Folic acid fortification, folate status and plasma homocysteine. *J Nutr.* 2002;132(8 Suppl):2466S-2470S. ([PubMed](#))
78. Homocysteine Lowering Trialists' Collaboration. Dose-dependent effects of folic acid on blood concentrations of homocysteine: a meta-analysis of the randomized trials. *Am J Clin Nutr.* 2005;82(4):806-812. ([PubMed](#))
79. Malinow MR, Bostom AG, Krauss RM. Homocyst(e)ine, diet, and cardiovascular diseases: a statement for healthcare professionals from the Nutrition Committee, American Heart Association. *Circulation.* 1999;99(1):178-182. ([PubMed](#))
80. van Meurs JB, Pare G, Schwartz SM, et al. Common genetic loci influencing plasma homocysteine concentrations and their effect on risk of coronary artery disease. *Am J Clin Nutr.* 2013;98(3):668-676. ([PubMed](#))
81. Holmes MV, Newcombe P, Hubacek JA, et al. Effect modification by population dietary folate on the association between MTHFR genotype, homocysteine, and stroke risk: a meta-analysis of genetic studies and randomised trials. *Lancet.* 2011;378(9791):584-594. ([PubMed](#))
82. Clarke R, Bennett DA, Parish S, et al. Homocysteine and coronary heart disease: meta-analysis of MTHFR case-control studies, avoiding publication bias. *PLoS Med.* 2012;9(2):e1001177. ([PubMed](#))
83. Ji Y, Tan S, Xu Y, et al. Vitamin B supplementation, homocysteine levels, and the risk of cerebrovascular disease: A meta-analysis. *Neurology.* 2013;81(15):1298-1307. ([PubMed](#))
84. Zhang C, Chi FL, Xie TH, Zhou YH. Effect of B-vitamin supplementation on stroke: a meta-analysis of randomized controlled trials. *PLoS One.* 2013;8(11):e81577. ([PubMed](#))
85. Mosca L, Benjamin EJ, Berra K, et al. Effectiveness-based guidelines for the prevention of cardiovascular disease in women--2011 update: a guideline from the American Heart Association. *J Am Coll Cardiol.* 2011;57(12):1404-1423. ([PubMed](#))
86. Wang X, Qin X, Demirtas H, et al. Efficacy of folic acid supplementation in stroke prevention: a meta-analysis. *Lancet.* 2007;369(9576):1876-1882. ([PubMed](#))

87. Yang Q, Botto LD, Erickson JD, et al. Improvement in stroke mortality in Canada and the United States, 1990 to 2002. *Circulation*. 2006;113(10):1335-1343. ([PubMed](#))
88. Lorenz MW, Markus HS, Bots ML, Rosvall M, Sitzer M. Prediction of clinical cardiovascular events with carotid intima-media thickness: a systematic review and meta-analysis. *Circulation*. 2007;115(4):459-467. ([PubMed](#))
89. Qin X, Xu M, Zhang Y, et al. Effect of folic acid supplementation on the progression of carotid intima-media thickness: a meta-analysis of randomized controlled trials. *Atherosclerosis*. 2012;222(2):307-313. ([PubMed](#))
90. de Bree A, van Mierlo LA, Draijer R. Folic acid improves vascular reactivity in humans: a meta-analysis of randomized controlled trials. *Am J Clin Nutr*. 2007;86(3):610-617. ([PubMed](#))
91. McNeil CJ, Beattie JH, Gordon MJ, Pirie LP, Duthie SJ. Nutritional B vitamin deficiency disrupts lipid metabolism causing accumulation of proatherogenic lipoproteins in the aorta adventitia of ApoE null mice. *Mol Nutr Food Res*. 2012;56(7):1122-1130. ([PubMed](#))
92. Blount BC, Mack MM, Wehr CM, et al. Folate deficiency causes uracil misincorporation into human DNA and chromosome breakage: implications for cancer and neuronal damage. *Proc Natl Acad Sci U S A*. 1997;94(7):3290-3295. ([PubMed](#))
93. Narayanan S, McConnell J, Little J, et al. Associations between two common variants C677T and A1298C in the methylenetetrahydrofolate reductase gene and measures of folate metabolism and DNA stability (strand breaks, misincorporated uracil, and DNA methylation status) in human lymphocytes in vivo. *Cancer Epidemiol Biomarkers Prev*. 2004;13(9):1436-1443. ([PubMed](#))
94. Rahman L, Voeller D, Rahman M, et al. Thymidylate synthase as an oncogene: a novel role for an essential DNA synthesis enzyme. *Cancer Cell*. 2004;5(4):341-351. ([PubMed](#))
95. Hubner RA, Liu JF, Sellick GS, Logan RF, Houlston RS, Muir KR. Thymidylate synthase polymorphisms, folate and B-vitamin intake, and risk of colorectal adenoma. *Br J Cancer*. 2007;97(10):1449-1456. ([PubMed](#))
96. Desmoulin SK, Wang L, Polin L, et al. Functional loss of the reduced folate carrier enhances the antitumor activities of novel antifolates with selective uptake by the proton-coupled folate transporter. *Mol Pharmacol*. 2012;82(4):591-600. ([PubMed](#))
97. Crider KS, Yang TP, Berry RJ, Bailey LB. Folate and DNA methylation: a review of molecular mechanisms and the evidence for folate's role. *Adv Nutr*. 2012;3(1):21-38. ([PubMed](#))
98. Butrum RR, Clifford CK, Lanza E. NCI dietary guidelines: rationale. *Am J Clin Nutr*. 1988;48(3 Suppl):888-895. ([PubMed](#))
99. Crider KS, Bailey LB, Berry RJ. Folic acid food fortification-its history, effect, concerns, and future directions. *Nutrients*. 2011;3(3):370-384. ([PubMed](#))
100. Qin X, Cui Y, Shen L, et al. Folic acid supplementation and cancer risk: A meta-analysis of randomized controlled trials. *Int J Cancer*. 2013;133(5):1033-1041. ([PubMed](#))
101. Vollset SE, Clarke R, Lewington S, et al. Effects of folic acid supplementation on overall and site-specific cancer incidence during the randomised trials: meta-analyses of data on 50,000 individuals. *Lancet*. 2013;381(9871):1029-1036. ([PubMed](#))
102. Kim DH, Smith-Warner SA, Spiegelman D, et al. Pooled analyses of 13 prospective cohort studies on folate intake and colon cancer. *Cancer Causes Control*. 2010;21(11):1919-1930. ([PubMed](#))

103. Gibson TM, Weinstein SJ, Pfeiffer RM, et al. Pre- and postfortification intake of folate and risk of colorectal cancer in a large prospective cohort study in the United States. *Am J Clin Nutr.* 2011;94(4):1053-1062. ([PubMed](#))
104. Stevens VL, McCullough ML, Sun J, Jacobs EJ, Campbell PT, Gapstur SM. High levels of folate from supplements and fortification are not associated with increased risk of colorectal cancer. *Gastroenterology.* 2011;141(1):98-105, 105 e101. ([PubMed](#))
105. Zschabitz S, Cheng TY, Neuhauser ML, et al. B vitamin intakes and incidence of colorectal cancer: results from the Women's Health Initiative Observational Study cohort. *Am J Clin Nutr.* 2013;97(2):332-343. ([PubMed](#))
106. Keum N, Giovannucci EL. Folic acid fortification and colorectal cancer risk. *Am J Prev Med.* 2014;46(3 Suppl 1):S65-72. ([PubMed](#))
107. Kennedy DA, Stern SJ, Moretti M, et al. Folate intake and the risk of colorectal cancer: a systematic review and meta-analysis. *Cancer Epidemiol.* 2011;35(1):2-10. ([PubMed](#))
108. Kim YI. Folate: a magic bullet or a double edged sword for colorectal cancer prevention? *Gut.* 2006;55(10):1387-1389. ([PubMed](#))
109. Paspatis GA, Kalafatis E, Oros L, Xourgias V, Koutsidou P, Karamanolis DG. Folate status and adenomatous colonic polyps. A colonoscopically controlled study. *Dis Colon Rectum.* 1995;38(1):64-67; discussion 67-68. ([PubMed](#))
110. Jaszewski R, Misra S, Tobi M, et al. Folic acid supplementation inhibits recurrence of colorectal adenomas: a randomized chemoprevention trial. *World J Gastroenterol.* 2008;14(28):4492-4498. ([PubMed](#))
111. Wu K, Platz EA, Willett WC, et al. A randomized trial on folic acid supplementation and risk of recurrent colorectal adenoma. *Am J Clin Nutr.* 2009;90(6):1623-1631. ([PubMed](#))
112. Logan RF, Grainge MJ, Shepherd VC, Armitage NC, Muir KR. Aspirin and folic acid for the prevention of recurrent colorectal adenomas. *Gastroenterology.* 2008;134(1):29-38. ([PubMed](#))
113. Figueiredo JC, Mott LA, Giovannucci E, et al. Folic acid and prevention of colorectal adenomas: a combined analysis of randomized clinical trials. *Int J Cancer.* 2011;129(1):192-203. ([PubMed](#))
114. Kennedy DA, Stern SJ, Matok I, et al. Folate intake, MTHFR polymorphisms, and the risk of colorectal cancer: a systematic review and meta-analysis. *J Cancer Epidemiol.* 2012;2012:952508. ([PubMed](#))
115. Ding W, Zhou DL, Jiang X, Lu LS. Methionine synthase A2756G polymorphism and risk of colorectal adenoma and cancer: evidence based on 27 studies. *PLoS One.* 2013;8(4):e60508. ([PubMed](#))
116. Nan H, Lee JE, Rimm EB, Fuchs CS, Giovannucci EL, Cho E. Prospective study of alcohol consumption and the risk of colorectal cancer before and after folic acid fortification in the United States. *Ann Epidemiol.* 2013;23(9):558-563. ([PubMed](#))
117. Slattery ML, Potter JD, Samowitz W, Schaffer D, Leppert M. Methylene tetrahydrofolate reductase, diet, and risk of colon cancer. *Cancer Epidemiol Biomarkers Prev.* 1999;8(6):513-518. ([PubMed](#))
118. Ma J, Stampfer MJ, Giovannucci E, et al. Methylene tetrahydrofolate reductase polymorphism, dietary interactions, and risk of colorectal cancer. *Cancer Res.* 1997;57(6):1098-1102. ([PubMed](#))
119. Larsson SC, Giovannucci E, Wolk A. Folate and risk of breast cancer: a meta-analysis. *J Natl Cancer Inst.* 2007;99(1):64-76. ([PubMed](#))

120. Liu M, Cui LH, Ma AG, Li N, Piao JM. Lack of effects of dietary folate intake on risk of breast cancer: an updated meta-analysis of prospective studies. *Asian Pac J Cancer Prev*. 2014;15(5):2323-2328. ([PubMed](#))
121. Brooks PJ, Zakhari S. Moderate alcohol consumption and breast cancer in women: from epidemiology to mechanisms and interventions. *Alcohol Clin Exp Res*. 2013;37(1):23-30. ([PubMed](#))
122. Rohan TE, Jain MG, Howe GR, Miller AB. Dietary folate consumption and breast cancer risk. *J Natl Cancer Inst*. 2000;92(3):266-269. ([PubMed](#))
123. Sellers TA, Kushi LH, Cerhan JR, et al. Dietary folate intake, alcohol, and risk of breast cancer in a prospective study of postmenopausal women. *Epidemiology*. 2001;12(4):420-428. ([PubMed](#))
124. Zhang S, Hunter DJ, Hankinson SE, et al. A prospective study of folate intake and the risk of breast cancer. *JAMA*. 1999;281(17):1632-1637. ([PubMed](#))
125. Tjonneland A, Christensen J, Olsen A, et al. Alcohol intake and breast cancer risk: the European Prospective Investigation into Cancer and Nutrition (EPIC). *Cancer Causes Control*. 2007;18(4):361-373. ([PubMed](#))
126. Bassett JK, Baglietto L, Hodge AM, et al. Dietary intake of B vitamins and methionine and breast cancer risk. *Cancer Causes Control*. 2013;24(8):1555-1563. ([PubMed](#))
127. Wang J, Wang B, Bi J, Di J. The association between two polymorphisms in the TYMS gene and breast cancer risk: a meta-analysis. *Breast Cancer Res Treat*. 2011;128(1):203-209. ([PubMed](#))
128. Weiner AS, Boyarskikh UA, Voronina EN, et al. Polymorphisms in the folate-metabolizing genes MTR, MTRR, and CBS and breast cancer risk. *Cancer Epidemiol*. 2012;36(2):e95-e100. ([PubMed](#))
129. Linabery AM, Johnson KJ, Ross JA. Childhood cancer incidence trends in association with US folic acid fortification (1986-2008). *Pediatrics*. 2012;129(6):1125-1133. ([PubMed](#))
130. Milne E, Royle JA, Miller M, et al. Maternal folate and other vitamin supplementation during pregnancy and risk of acute lymphoblastic leukemia in the offspring. *Int J Cancer*. 2010;126(11):2690-2699. ([PubMed](#))
131. Yan J, Yin M, Dreyer ZE, et al. A meta-analysis of MTHFR C677T and A1298C polymorphisms and risk of acute lymphoblastic leukemia in children. *Pediatr Blood Cancer*. 2012;58(4):513-518. ([PubMed](#))
132. Alzheimer's Association. 2013 Alzheimer's Disease Fact and Figures. *Alzheimer's & Dementia*. 9(2). Available at: [http://www.alz.org/downloads/facts\\_figures\\_2013.pdf](http://www.alz.org/downloads/facts_figures_2013.pdf). Accessed 9/9/13.
133. Hughes TF, Andel R, Small BJ, et al. Midlife fruit and vegetable consumption and risk of dementia in later life in Swedish twins. *Am J Geriatr Psychiatry*. 2010;18(5):413-420. ([PubMed](#))
134. Weir DG, Scott JM. Brain function in the elderly: role of vitamin B12 and folate. *Br Med Bull*. 1999;55(3):669-682. ([PubMed](#))
135. Faux NG, Ellis KA, Porter L, et al. Homocysteine, vitamin B12, and folic acid levels in Alzheimer's disease, mild cognitive impairment, and healthy elderly: baseline characteristics in subjects of the Australian Imaging Biomarker Lifestyle study. *J Alzheimers Dis*. 2011;27(4):909-922. ([PubMed](#))
136. Van Dam F, Van Gool WA. Hyperhomocysteinemia and Alzheimer's disease: A systematic review. *Arch Gerontol Geriatr*. 2009;48(3):425-430. ([PubMed](#))
137. Morris MC, Evans DA, Bienias JL, et al. Dietary folate and vitamin B12 intake and cognitive decline among community-dwelling older persons. *Arch Neurol*. 2005;62(4):641-645. ([PubMed](#))

138. Morris MC, Evans DA, Schneider JA, Tangney CC, Bienias JL, Aggarwal NT. Dietary folate and vitamins B-12 and B-6 not associated with incident Alzheimer's disease. *J Alzheimers Dis.* 2006;9(4):435-443. ([PubMed](#))
139. Wald DS, Kasturiratne A, Simmonds M. Serum homocysteine and dementia: meta-analysis of eight cohort studies including 8669 participants. *Alzheimers Dement.* 2011;7(4):412-417. ([PubMed](#))
140. Ho RC, Cheung MW, Fu E, et al. Is high homocysteine level a risk factor for cognitive decline in elderly? A systematic review, meta-analysis, and meta-regression. *Am J Geriatr Psychiatry.* 2011;19(7):607-617. ([PubMed](#))
141. Nilforooshan R, Broadbent D, Weaving G, et al. Homocysteine in Alzheimer's disease: role of dietary folate, vitamin B6 and B12. *Int J Geriatr Psychiatry.* 2011;26(8):876-877. ([PubMed](#))
142. Wald DS, Kasturiratne A, Simmonds M. Effect of folic acid, with or without other B vitamins, on cognitive decline: meta-analysis of randomized trials. *Am J Med.* 2010;123(6):522-527 e522. ([PubMed](#))
143. Ford AH, Almeida OP. Effect of homocysteine lowering treatment on cognitive function: a systematic review and meta-analysis of randomized controlled trials. *J Alzheimers Dis.* 2012;29(1):133-149. ([PubMed](#))
144. Nachum-Biala Y, Troen AM. B-vitamins for neuroprotection: narrowing the evidence gap. *Biofactors.* 2012;38(2):145-150. ([PubMed](#))
145. Smith AD, Smith SM, de Jager CA, et al. Homocysteine-lowering by B vitamins slows the rate of accelerated brain atrophy in mild cognitive impairment: a randomized controlled trial. *PLoS One.* 2010;5(9):e12244. ([PubMed](#))
146. Douaud G, Refsum H, de Jager CA, et al. Preventing Alzheimer's disease-related gray matter atrophy by B-vitamin treatment. *Proc Natl Acad Sci U S A.* 2013;110(23):9523-9528. ([PubMed](#))
147. Watkins D, Rosenblatt DS. Update and new concepts in vitamin responsive disorders of folate transport and metabolism. *J Inherit Metab Dis.* 2012;35(4):665-670. ([PubMed](#))
148. Zhao R, Min SH, Qiu A, et al. The spectrum of mutations in the PCFT gene, coding for an intestinal folate transporter, that are the basis for hereditary folate malabsorption. *Blood.* 2007;110(4):1147-1152. ([PubMed](#))
149. Borzutzky A, Crompton B, Bergmann AK, et al. Reversible severe combined immunodeficiency phenotype secondary to a mutation of the proton-coupled folate transporter. *Clin Immunol.* 2009;133(3):287-294. ([PubMed](#))
150. Sofer Y, Harel L, Sharkia M, Amir J, Schoenfeld T, Straussberg R. Neurological manifestations of folate transport defect: case report and review of the literature. *J Child Neurol.* 2007;22(6):783-786. ([PubMed](#))
151. Diop-Bove N, Kronn D, Goldman ID. Hereditary folate malabsorption. In: Pagon RA, Adam MP, Bird TD, Dolan CR, Fong CT, Stephens K, eds. *GeneReviews™* [Internet]. Seattle, WA: University of Washington, Seattle; 2008. ([PubMed](#))
152. Frye RE, Sequeira JM, Quadros EV, James SJ, Rossignol DA. Cerebral folate receptor autoantibodies in autism spectrum disorder. *Mol Psychiatry.* 2013;18(3):369-381. ([PubMed](#))
153. Grapp M, Just IA, Linnankivi T, et al. Molecular characterization of folate receptor 1 mutations delineates cerebral folate transport deficiency. *Brain.* 2012;135(Pt 7):2022-2031. ([PubMed](#))
154. Ramaekers VT, Blau N, Sequeira JM, Nassogne MC, Quadros EV. Folate receptor autoimmunity and cerebral folate deficiency in low-functioning autism with neurological deficits. *Neuropediatrics.*

2007;38(6):276-281. ([PubMed](#))

155. Ramaekers VT, Hausler M, Opladen T, Heimann G, Blau N. Psychomotor retardation, spastic paraplegia, cerebellar ataxia and dyskinesia associated with low 5-methyltetrahydrofolate in cerebrospinal fluid: a novel neurometabolic condition responding to folinic acid substitution. *Neuropediatrics*. 2002;33(6):301-308. ([PubMed](#))

156. Banka S, Blom HJ, Walter J, et al. Identification and characterization of an inborn error of metabolism caused by dihydrofolate reductase deficiency. *Am J Hum Genet*. 2011;88(2):216-225. ([PubMed](#))

157. Cario H, Smith DE, Blom H, et al. Dihydrofolate reductase deficiency due to a homozygous DHFR mutation causes megaloblastic anemia and cerebral folate deficiency leading to severe neurologic disease. *Am J Hum Genet*. 2011;88(2):226-231. ([PubMed](#))

158. Pfeiffer CM, Hughes JP, Lacher DA, et al. Estimation of trends in serum and RBC folate in the US population from pre- to postfortification using assay-adjusted data from the NHANES 1988-2010. *J Nutr*. 2012;142(5):886-893. ([PubMed](#))

159. Folate. In: Hendler SS, Rorvik, D.R., ed. *PDR for Nutritional Supplements*. 2nd ed. Montvale: Physicians' Desk Reference Inc.; 2008.

160. Wiesinger H, Eydeler U, Richard F, et al. Bioequivalence evaluation of a folate-supplemented oral contraceptive containing ethinylestradiol/drospirenone/levomefolate calcium versus ethinylestradiol/drospirenone and levomefolate calcium alone. *Clin Drug Investig*. 2012;32(10):673-684. ([PubMed](#))

161. Tinker SC, Cogswell ME, Devine O, Berry RJ. Folic acid intake among US women aged 15-44 years, National Health and Nutrition Examination Survey, 2003-2006. *Am J Prev Med*. 2010;38(5):534-542. ([PubMed](#))

162. Kelly P, McPartlin J, Goggins M, Weir DG, Scott JM. Unmetabolized folic acid in serum: acute studies in subjects consuming fortified food and supplements. *Am J Clin Nutr*. 1997;65(6):1790-1795. ([PubMed](#))

163. Morris MS, Jacques PF, Rosenberg IH, Selhub J. Folate and vitamin B-12 status in relation to anemia, macrocytosis, and cognitive impairment in older Americans in the age of folic acid fortification. *Am J Clin Nutr*. 2007;85(1):193-200. ([PubMed](#))

164. Morris MS, Jacques PF, Rosenberg IH, Selhub J. Circulating unmetabolized folic acid and 5-methyltetrahydrofolate in relation to anemia, macrocytosis, and cognitive test performance in American seniors. *Am J Clin Nutr*. 2010;91(6):1733-1744. ([PubMed](#))

165. Troen AM, Mitchell B, Sorensen B, et al. Unmetabolized folic acid in plasma is associated with reduced natural killer cell cytotoxicity among postmenopausal women. *J Nutr*. 2006;136(1):189-194. ([PubMed](#))

166. Tam C, O'Connor D, Koren G. Circulating unmetabolized folic acid: relationship to folate status and effect of supplementation. *Obstet Gynecol Int*. 2012;2012:485179. ([PubMed](#))

167. Apeland T, Mansoor MA, Strandjord RE. Antiepileptic drugs as independent predictors of plasma total homocysteine levels. *Epilepsy Res*. 2001;47(1-2):27-35. ([PubMed](#))

168. Wilson SM, Bivins BN, Russell KA, Bailey LB. Oral contraceptive use: impact on folate, vitamin B(6), and vitamin B(1)(2) status. *Nutr Rev*. 2011;69(10):572-583. ([PubMed](#))